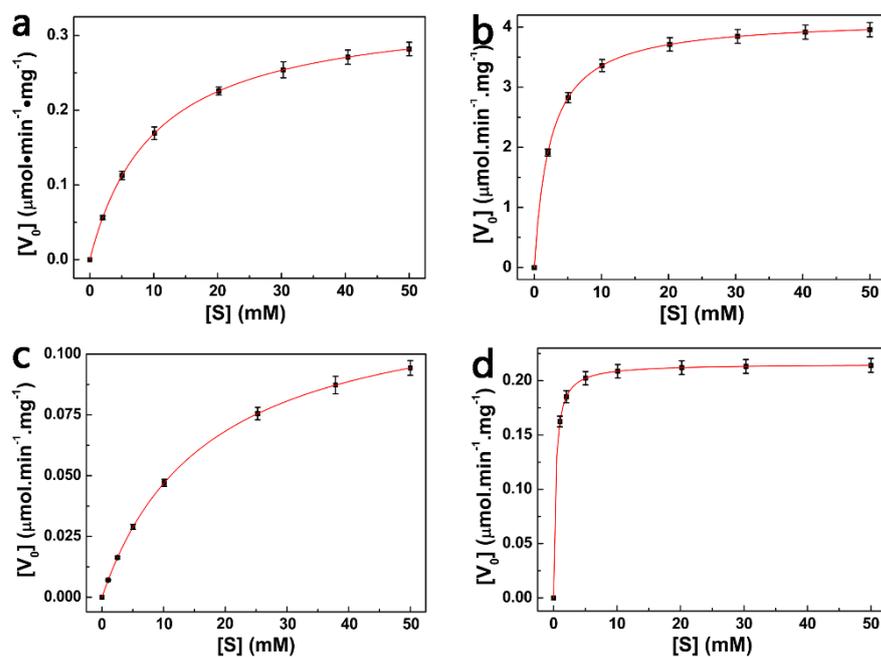
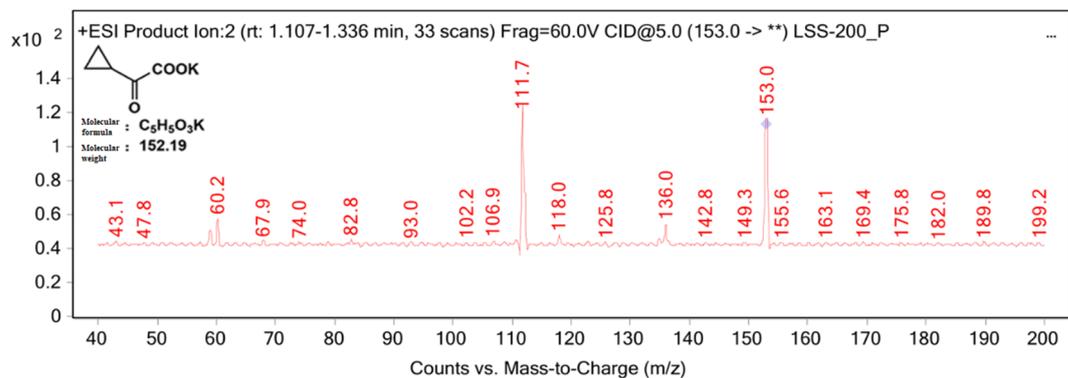


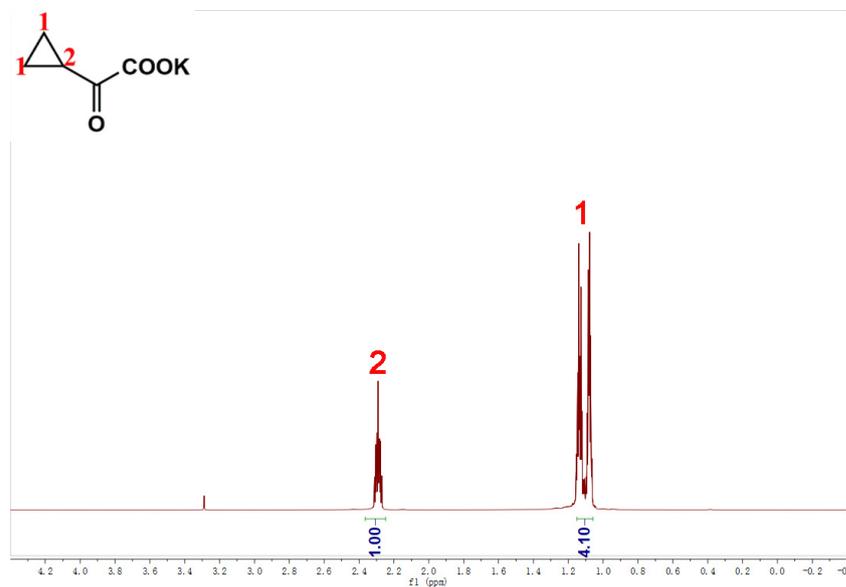
**Figure S1.** Asymmetric synthesis of (S)-cyclopropylglycine catalyzed by whole-cell *E. coli* (KLT) (a), a combination of the two native enzymes *E. coli* (Ti-LDH) and *E. coli* (Kp-FDH) (b), and *E. coli* (TLK) (c). The reaction was performed in phosphate-buffered saline (PBS, pH8.0) containing  $\text{Na}_2\text{HPO}_4$ ,  $\text{KH}_2\text{PO}_4$ , NaCl and KCl with use of lyophilized cells (75 mg), potassium cyclopropyl-glyoxylate (4 mmol), ammonium formate (12 mmol), 0.6 mM NADH at total volume of 10 mL and 40 °C. pH was kept at 8.0-9.0 with NaOH.



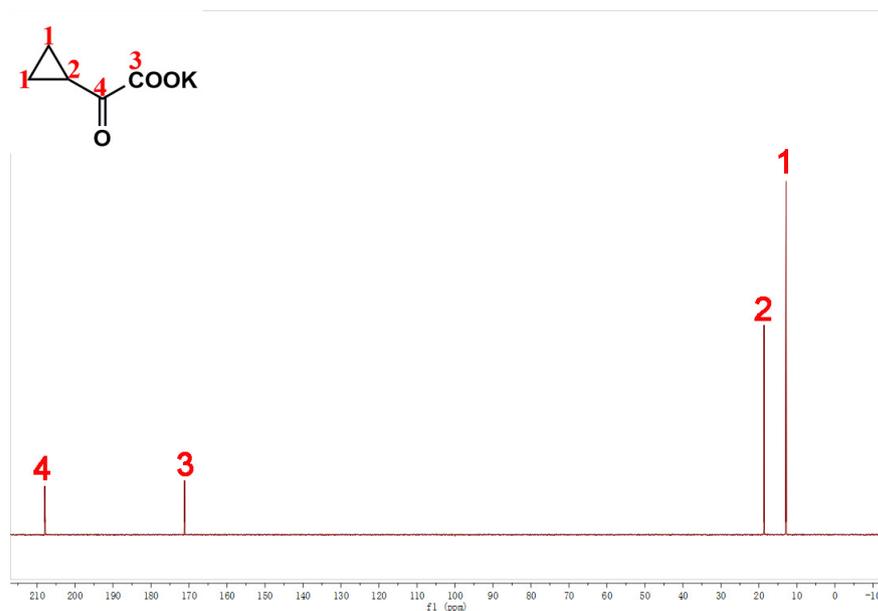
**Figure S2.** Steady-state kinetic of activities for different recombinant enzymes. a, c), Dependence of coenzyme regeneration activity of *Kp*-FDH and *Kp*-FDH domains in the fusion enzyme TLK on ammonium formate concentration, respectively. b, d), Dependence of reductive amination activity of *Ti*-LDH and *Ti*-LDH domains in the fusion enzyme TLK on potassium cyclopropylglyoxylate concentration, respectively. Analysis was done using Origin 2016.



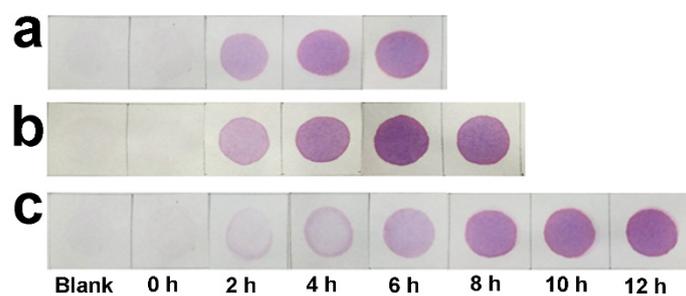
**Figure S3.** The LC-MS of potassium cyclopropylglyoxylate



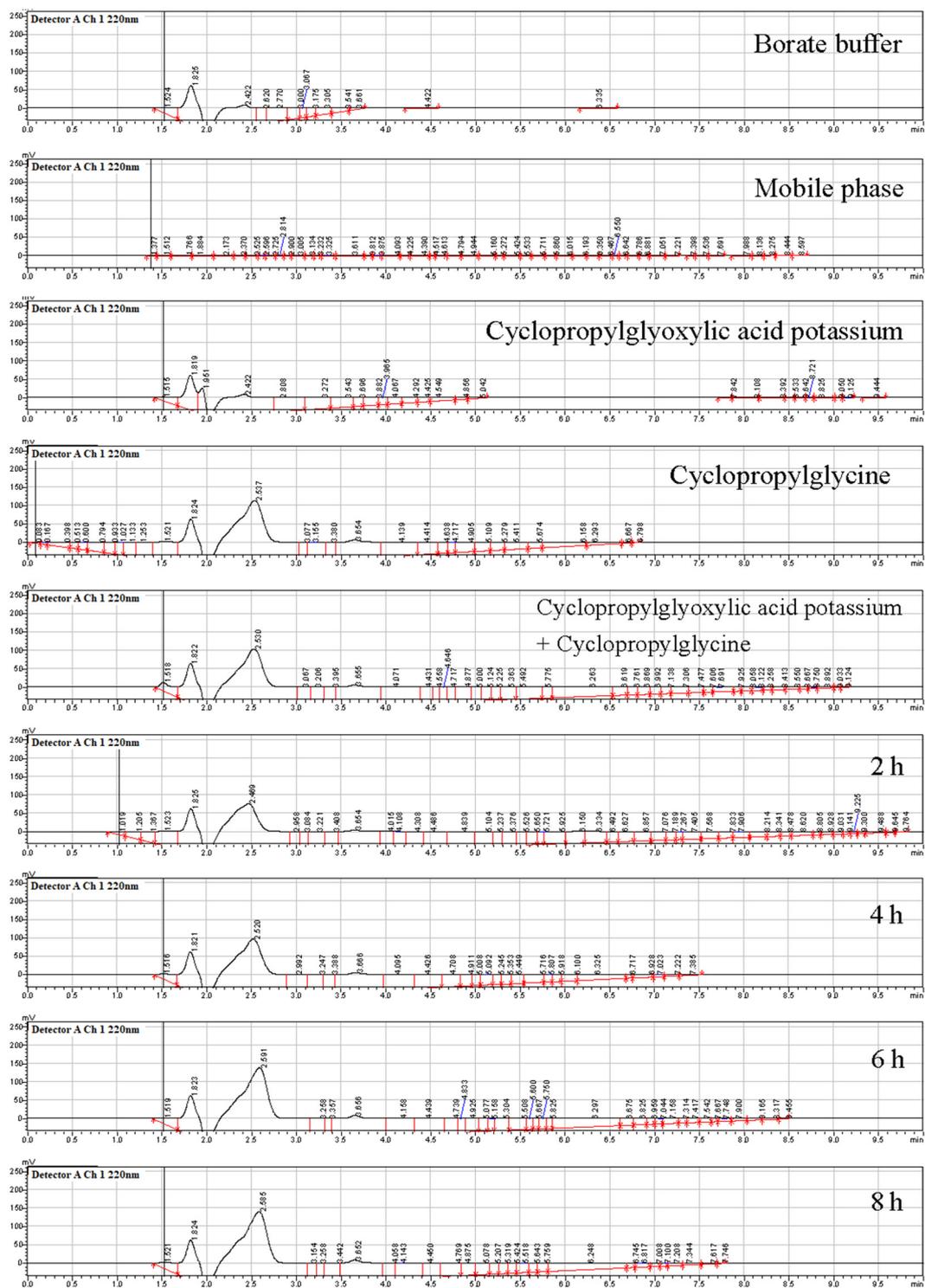
**Figure S4.** <sup>1</sup>H NMR of potassium cyclopropylglyoxylate (600 MHz, D<sub>2</sub>O).  $\delta = 1.0-1.2$  (4H, m),  $2.2-2.4$  (1H, m).



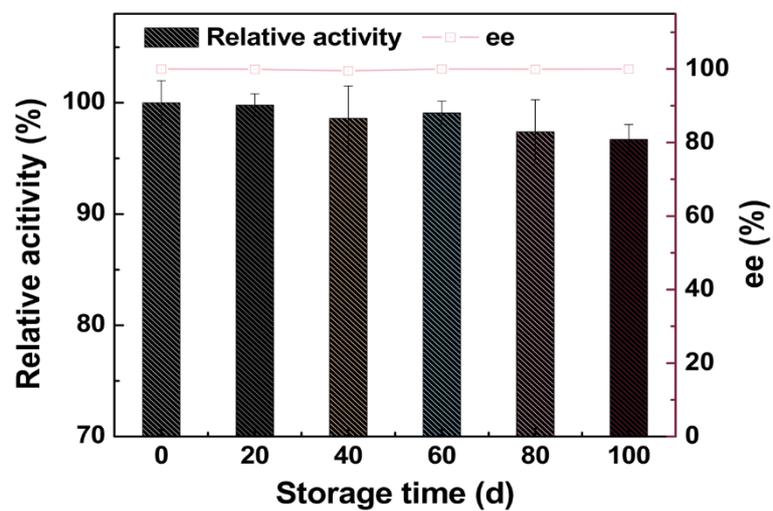
**Figure S5.**  $^{13}\text{C}$  NMR of potassium cyclopropylglyoxylate (600 MHz,  $\text{D}_2\text{O}$ ).  $\delta = 13.8$  ( $2\text{CH}_2$ ), 19.6 (CH), 172.3 (C), 209.1 (C).



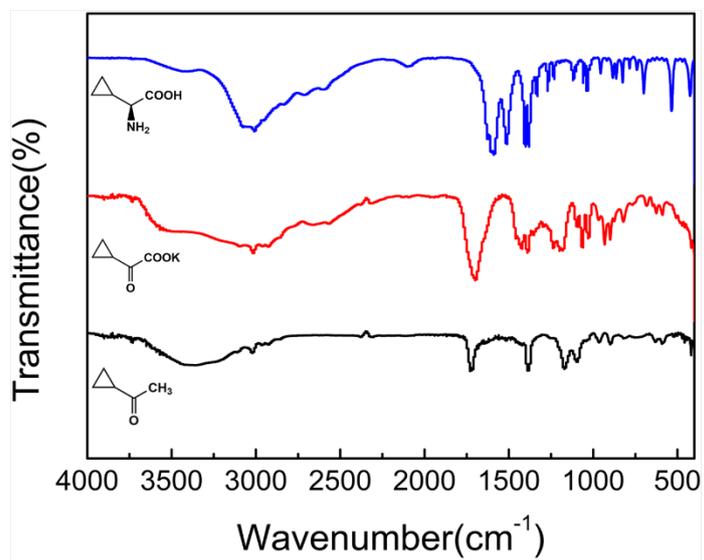
**Figure S6.** Asymmetric synthesis of (S)-cyclopropylglycine catalyzed by whole-cell *E. coli* (TLK) in the presence of 7.5 g/L biocatalysts and 0.6 mM cofactor at different concentration of potassium cyclopropylglyoxylate. (a), 100 g/L, (b) 120 g/L, and (c) 140 g/L. Other conditions: reaction medium was PBS (pH8.0), and pH was kept at 8.0-9.0 with NaOH and 40 °C.



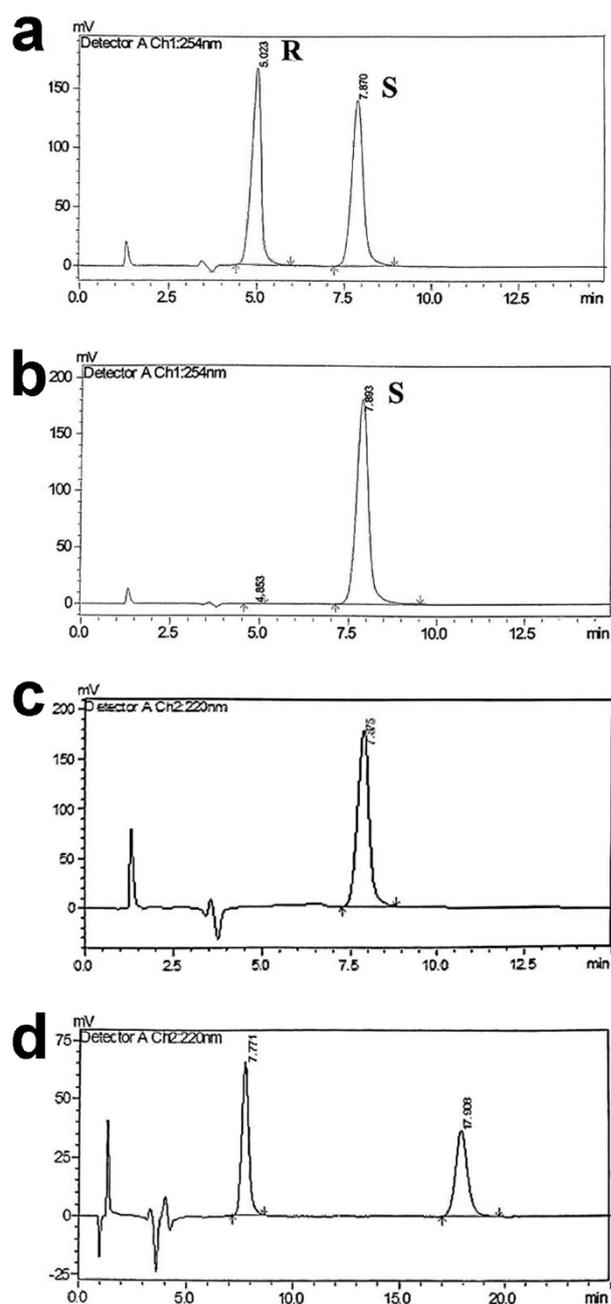
**Figure S7.** HPLC analysis of the generated (S)-cyclopropylglycine catalyzed by *E. coli* (KLT) at 120 g/L potassium cyclopropylglyoxylate in the presence of 7.5 g/L biocatalysts and 0.6 mM cofactor. Other conditions: reaction medium was PBS (pH8.0); pH was kept at 8.0-9.0 with NaOH at 40 °C.



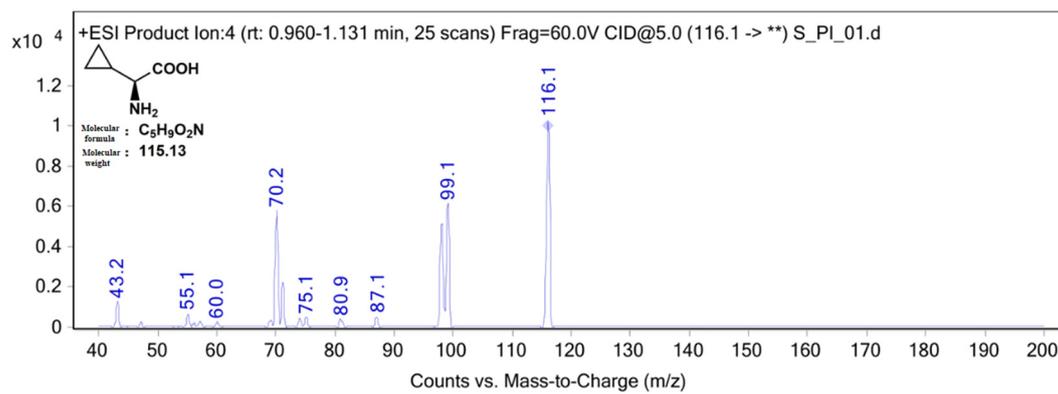
**Figure S8.** The influence of storage time on the biocatalytic activity of the bifunctional enzyme TLK. The biocatalytic process was conducted under standard conditions.



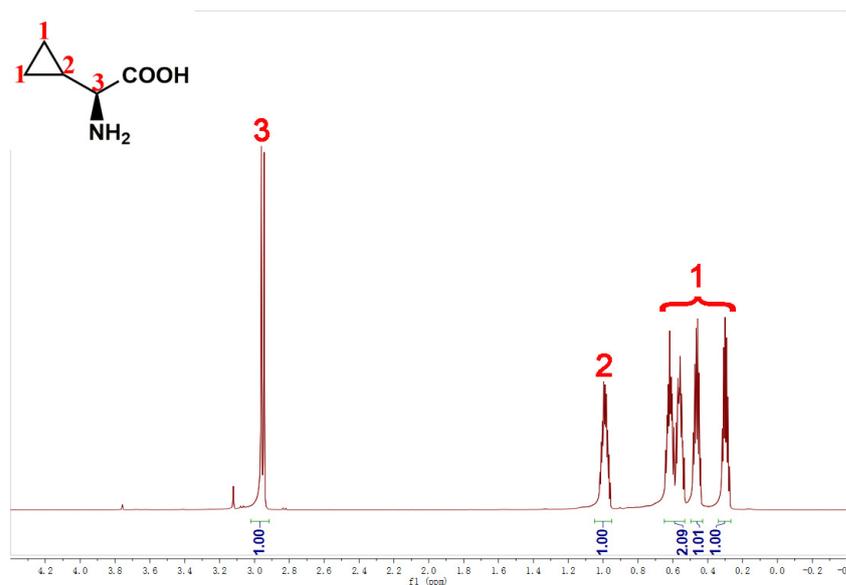
**Figure S9.** FT-IR spectra of cyclopropyl methyl ketone, potassium cyclopropylglyoxylate and (S)-cyclopropylglycine



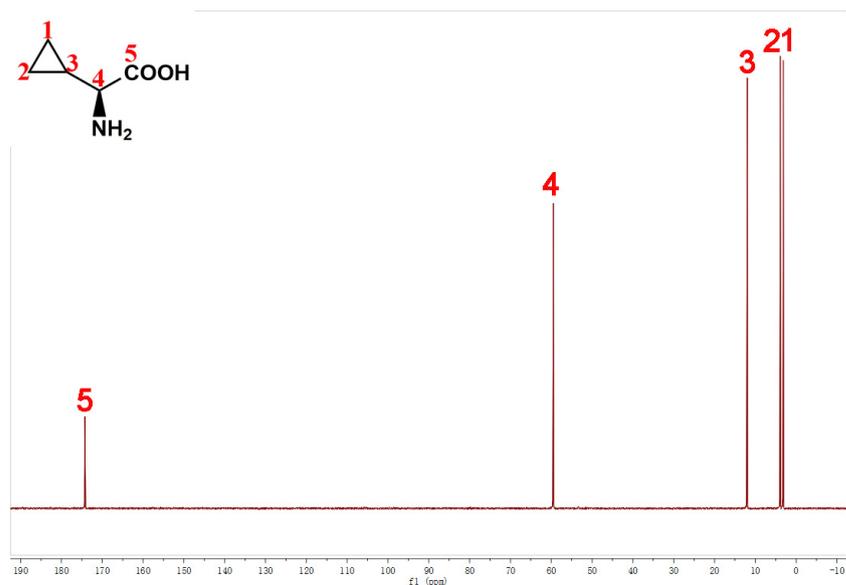
**Figure S10.** Determination of optical purity of the products by HPLC. a) The retention times for the standard (R/S)-cyclopropylglycine were 5.023 and 7.870 min. b) The retention time for the standard (S)-cyclopropylglycine was 7.893 min. c) The retention time for the synthesized (S)-cyclopropylglycine was 7.905 min. d) The collected reaction mixture supernatant after treated with trifluoroacetic acid and subsequent active carbon. The retention time for (S)-cyclopropylglycine and potassium cyclopropylglyoxylate was 7.771 min and 17.903 min, respectively.



**Figure S11.** The LC-MS of (S)-Cyclopropylglycine



**Figure S12.** <sup>1</sup>H NMR of (S)-Cyclopropylglycine (600 MHz, D<sub>2</sub>O).  $\delta = 0.2-0.3$  (1H, m),  $0.4-0.5$  (1H, m),  $0.5-0.7$  (2H, m),  $0.9-1.1$  (1H, m),  $2.9$ (1H, s).



**Figure S13.** <sup>13</sup>C NMR of (S)-Cyclopropylglycine (600 MHz, D<sub>2</sub>O).  $\delta = 4.4$  (CH<sub>2</sub>), 5.8 (CH<sub>2</sub>), 13.2 (CH), 59.2 (CH), 174.8 (C);

MKIFDYMEKYDYEQLVMCQDKESGLKAIICIHVTTLGPALGGMRMWTYASEEEAIEDALRLGRGMTYKN  
AAAGLNLGGGKTVIIGDPRKDKNEAMFRALGRFIQGLNGRYITAEDVGTTVEDMDIIHEETRYVTGVSPAF  
GSSGNPSPVTAYGVYRGMKAAAKEAFGDDSLEGKVVAVQGVGHVAYELCKHLHNEGAKLIVTDINKENA  
DRAVQEFGAEFVHPDKIYDVECDIFAPCALGAIINDETIERLKCKVVAGSANNQLKEERHGKMLEEKGIVY  
APDYVINAGGVINVADELLGYNRERAMKKVEGIYDKILKVFEIAKRDGIPSYLAADRMAEERIEMMRKTR  
STFLQDQRNLINFNNKGGGSMKIVLVLYSAGKHAADPKLYGCIENELGIRQWLEKGGHELVTTSDEK  
ENSELEKHIPDADVIISTPFHPAYITKERIQKAKKLLLVVAGVGS DHIDL DYIEQNGLDISVLEVTGSNVVS  
VAEHVVMTILNLVRNFVPAHEQIVNHGWDVAIAKDAYDIEGKTIATIGAGRIGYRVLERLVAFNPKELLY  
YDYQGLPKEAEEKVGARRVDTVEELVAQADVVTVNAPLHAGTKGLV NKELLSKFKKGAWLVNTARGAI  
CNAQDVADAVASGQLRGYGGDVWFPQPAPKDHPWRDMRNKYGYGNAMTPHYS GTTLDAQVRYAEGTK  
NILNSFLTKKFDYRPQDVILLNGKYKTKAYGNDKKVA

**Figure 14.** The amino acid sequence of TLK.