

*Supplemental Materials*

# **CRISPR-Cas9 KO Cell Line Generation and Development of a Cell-Based Potency Assay for rAAV-FKRP Gene Therapy**

**Marine Geoffroy<sup>1,2,\*</sup>, Louna Pili<sup>1,2</sup>, Valentina Buffa<sup>1,2</sup>, Maëlle Caroff<sup>1,2</sup>, Anne Bigot<sup>3</sup>, Evelyne Gicquel<sup>1,2</sup>, Grégory Rouby<sup>1,2</sup>, Isabelle Richard<sup>1,2,4</sup> and Romain Fragnoud<sup>1,2,\*</sup>**

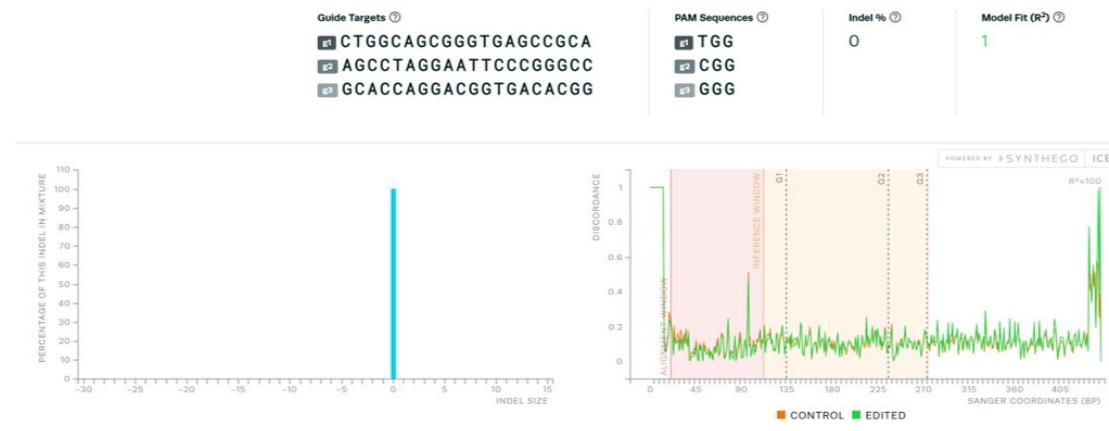
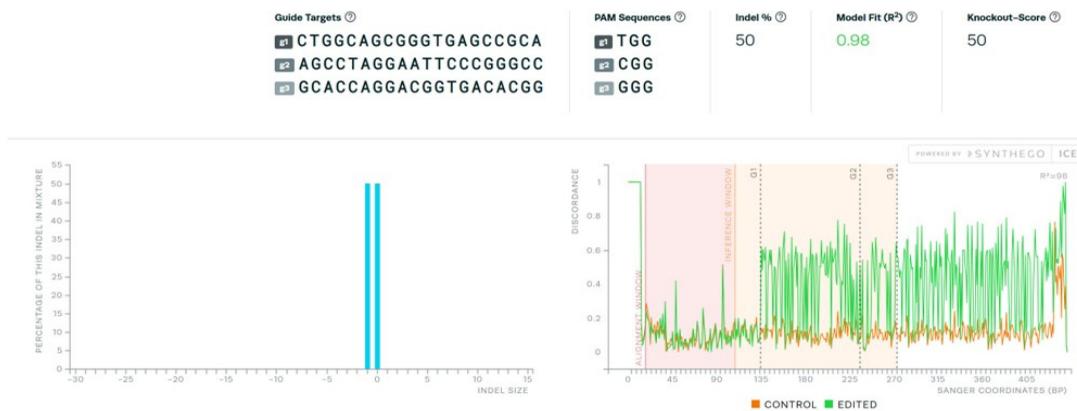
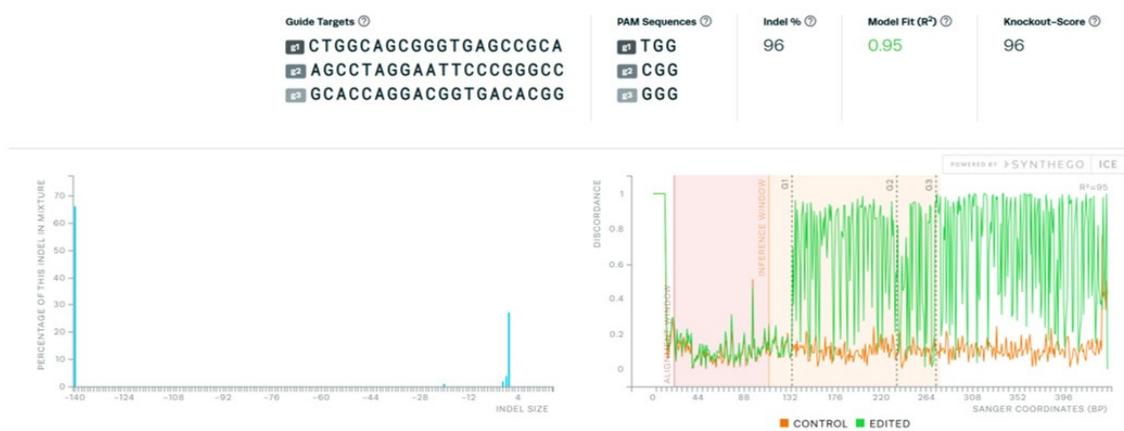
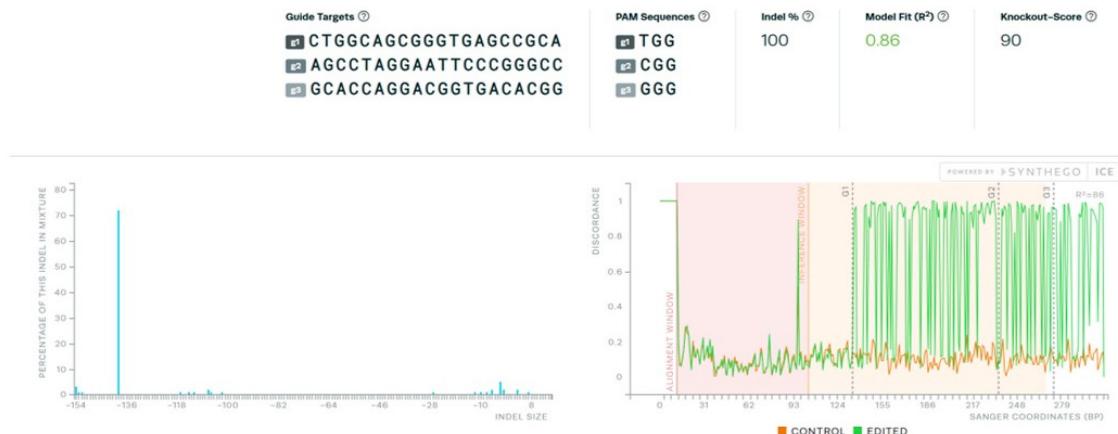
<sup>1</sup> Généthon, 91000 Evry-Courcouronnes, France

<sup>2</sup> Université Paris-Saclay/Université Evry, INSERM, Généthon, Integrare Research Unit, UMR\_S951, 91000 Evry, France

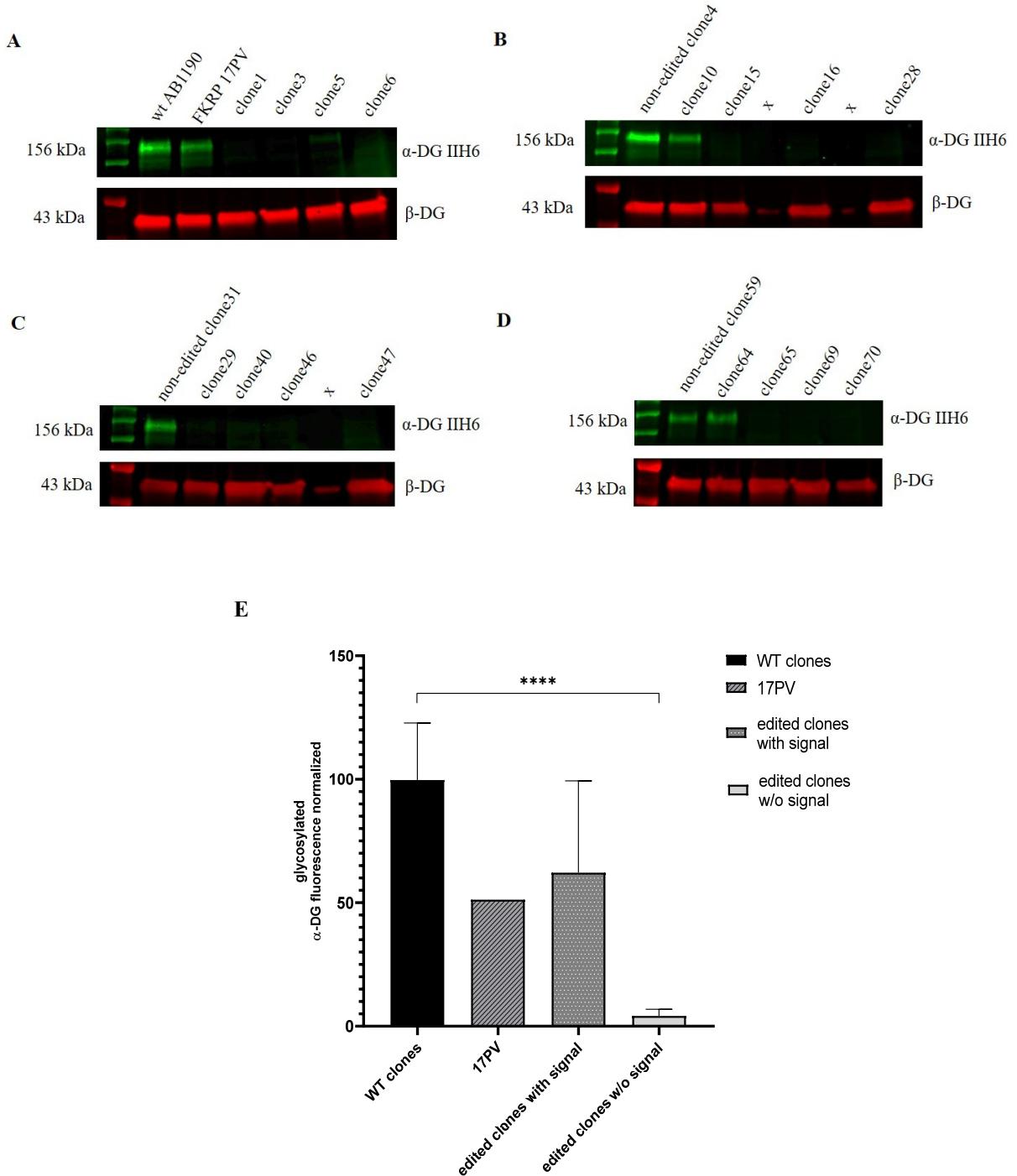
<sup>3</sup> Institut de Myologie, Université Pierre et Marie Curie Paris 6, UM76 Univ. Paris 6/U974 UMR7215, CNRS Pitié-Salpêtrière-INSERM, UMRS 974, 75000 Paris, France

<sup>4</sup> Atamyo Therapeutics, F-91000 Evry, France

\* Correspondence: marine.geoffroy@sqy-synthena.com (M.G.); rfragnoud@genethon.fr (R.F.); Tel.: +33-(0)169471042 (R.F.)

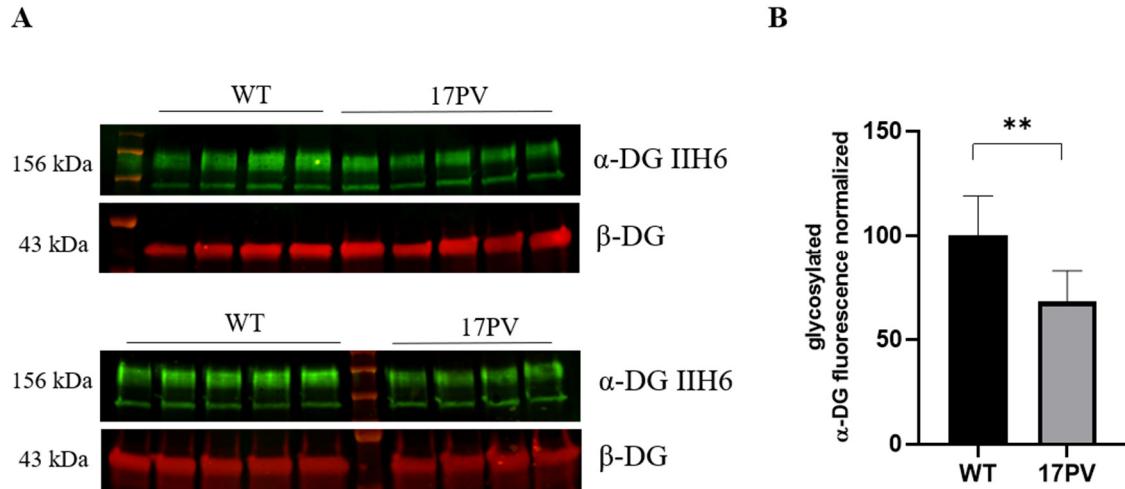
**A****B****C****D**

**Figure S1.** Gene editing efficiency and frequency of InDels determined using the ICE Software. **(A)** Representation of InDels in the non-edited wt clones (X/X); Top: sgRNAs sequences used and percentage of InDels for the mixture of sgRNAs. Bottom left: ICE InDels profiles generated by the sgRNAs. Bottom right: Comparison of the reference non-edited sequence (control in orange) and the non-edited wt clone (edited in green). **(B)** Representation of InDels in the edited heterozygous clones (X/A); Top: sgRNAs sequences used and percentage of InDels for the mixture of sgRNAs. Bottom left: ICE InDels profiles generated by the sgRNAs. Bottom right: Comparison of the reference non-edited sequence (control in orange) and the edited heterozygous clone (edited in green). **(C)** Representation of InDels in the edited compound heterozygous clones (A'/A); Top: sgRNAs sequences used and percentage of InDels for the mixture of sgRNAs. Bottom left: ICE InDels profiles generated by the sgRNAs. Bottom right: Comparison of the reference sequence non-edited (control in orange) and the edited compound heterozygous clone (edited in green). **(D)** Representation of InDels in the edited homozygous clones (A/A); Top: sgRNAs sequences used and percentage of InDels for the mixture of sgRNAs. Bottom left: ICE InDels profiles generated by the sgRNAs. Bottom right: Comparison of the reference non-edited sequence (control in orange) and the edited homozygous clone (edited in green).

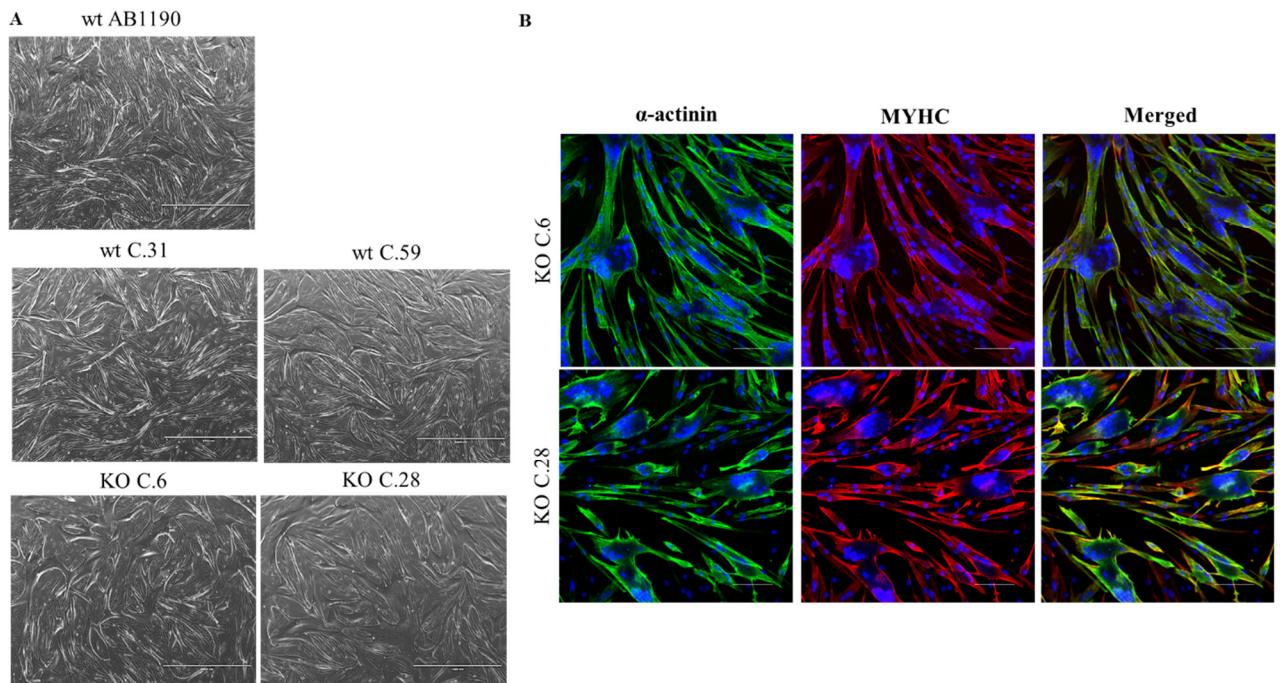


**Figure S2.** Expression of glycosylated  $\alpha$ -DG in the selection of 16 clones. **(A)** Expression of the glycosylated  $\alpha$ -DG using IIH6 antibody by western blot from the original wt myotubes (wt AB1190), the immortalized LGMDR9 patients myotubes (FKRP 17PV), and the edited clones (clone 1, clone 3, clone 5 and clone 6).  $\beta$ -DG was used as housekeeping protein. **(B)** Expression of the glycosylated  $\alpha$ -DG using IIH6 antibody by western blot from the non-edited clone 4 and the edited clones (clone 10, clone 15, clone 16 and clone 28).  $\beta$ -DG was used as housekeeping protein. The “x” corresponds to an empty well. **(C)** Expression of the glycosylated  $\alpha$ -DG using IIH6 antibody by western blot from the non-edited clone 31 and the edited clones (clone 29, clone 40, clone 46 and clone 47).  $\beta$ -DG was used as housekeeping protein. The “x” corresponds to an empty well. **(D)** Expression of the glycosylated  $\alpha$ -DG using IIH6 antibody by western blot from the non-edited clone 59 and the edited clones (clone 64, clone 65, clone 69 and clone 70).  $\beta$ -DG was used as housekeeping protein. **(E)**

Quantification of glycosylated  $\alpha$ -DG normalized in 4 groups: WT/non edited clone (AB1190/clone 4, 31 and 59), FKRP 17PV cell line, edited clones with glycosylated  $\alpha$ -DG signal (clones 5, 10 and 64), edited clones with no glycosylated  $\alpha$ -DG signal (clones 1, 3, 6, 15, 16, 28, 29, 40, 47, 65, 69 and 70). \*\*\* p-value < 0.0001 following an unpaired t-test.



**Figure S3.** Expression of glycosylated  $\alpha$ -DG in WT and FKRP immortalized cell lines. **(A)** Expression of the glycosylated  $\alpha$ -DG using IIH6 antibody by western blot from the original wt myotubes (wt AB1190) and the immortalized LGMDR9 patients myotubes (FKRP 17PV). Two different experiments were performed. **(B)** Quantification and normalization of glycosylated  $\alpha$ -DG normalized (n=9 per cell lines). \*\* p-value = 0.0011 following an unpaired t-test.



**Figure S4.** Myotube differentiation observed by microscopic analysis and immunofluorescence staining in edited clones. **(A)** Brightfield pictures of the original wt cells (AB1190), the non-edited wt clones (wt C.31 and wt C.59) and the edited wt clones (KO C.6 and KO C.28) at myotube stage. Scale bars = 1,000  $\mu$ m. **(B)** Immunofluorescence staining ( $\alpha$ -actinin and MYHC) of the edited clones (KO C.6 and KO C.28). Cell nuclei were labelled with DAPI dye. Scale bars = 100  $\mu$ m.

**Table S1.** *In silico* analysis of off-targets with CRISPOR TEFOR **(A)** Top 10 locus of off-targets for sgRNA1. **(B)** Top 10 locus of off-targets for sgRNA2. **(C)** Top 10 locus of off-targets for sgRNA3.

A	Locus	mismatchCount	cfdOfftargetScore	chrom	start	end	strand
	<b>FKRP</b>	<b>0</b>	<b>100</b>	<b>chr19</b>	<b>46755455</b>	<b>46755477</b>	-
	intergenic:RP11-1151B14.2-RP11-1151B14.3	4	0.51	chr18	56112872	56112894	-
	intergenic:AL136967.1-RP11-328M4.2	3	0.46	chr6	41384109	41384131	+
	intron:ZNF862	4	0.40	chr7	149551791	149551813	-
	intergenic:RP11-128M1.1-TGM3	3	0.34	chr20	2268800	2268822	+
	exon:DBNL/PGAM2/AC017116.11	4	0.34	chr7	44101156	44101178	-
	intergenic:LMBR1-RNF32	3	0.31	chr7	156462214	156462236	+
	intron:MYO18B	4	0.30	chr22	26145944	26145966	+
	intergenic:RP11-386G21.2-XKR4	4	0.30	chr8	56083410	56083432	+
	intergenic:RP11-761N21.1-RP11-520A21.1	4	0.29	chr3	40950300	40950322	-
	intergenic:RXRA-RP11-473E2.2	4	0.28	chr9	137393918	137393940	-

B	Locus	mismatchCount	cfdOfftargetScore	chrom	start	end	strand
	<b>FKRP</b>	<b>0</b>	<b>100</b>	<b>chr19</b>	<b>46755556</b>	<b>46755578</b>	+
	intergenic:CTC-461H2.2-RHPN2	4	0.71	chr19	33554035	33554057	+
	intergenic:KCNN3-PMVK	3	0.67	chr1	154845237	154845259	-
	lnc:AC006116.17/ZSCAN5A/Y RNA	3	0.62	chr19	56826681	56826703	+
	intergenic:RNU7-145P-RP11-142I20.1	4	0.53	chr18	38950440	38950462	-
	intergenic:RPS23P3-RNU6-699P	4	0.50	chr4	67430059	67430081	+
	lnc:RP11-50E11.2/RP11-50E11.3	4	0.41	chr10	52397448	52397470	-
	intron:DFNB31	4	0.40	chr9	117170569	117170591	-
	intergenic:COMP-UPF1	4	0.39	chr19	18924078	18924100	-
	exon:C6orf226	4	0.33	chr6	42858415	42858437	-
	exon:CD22	3	0.30	chr19	35820497	35820519	+

C	Locus	mismatchCount	cfdOfftargetScore	chrom	start	end	strand
	<b>FKRP</b>	<b>0</b>	<b>100</b>	<b>chr19</b>	<b>46755594</b>	<b>46755616</b>	-
	exon:PYGM	4	0.57	chr11	64518824	64518846	+
	intron:RP11-890B15.2	4	0.54	chr11	130723063	130723085	-
	intron:GPR56	4	0.39	chr16	57692790	57692812	+
	intergenic:SNAP25-AS1-SDAD1P2	4	0.37	chr20	10360573	10360595	+
	intron:AGAP3	3	0.33	chr7	150805390	150805412	-
	intergenic:MDH2-AC005077.14	4	0.32	chr7	75698884	75698906	+
	intron:CUX1	4	0.31	chr7	101818983	101819005	-
	intron:AC008271.1	4	0.31	chr2	15831614	15831636	-
	intergenic:RP11-324D17.2-RP11-324D17.1	4	0.30	chr16	54276113	54276135	-
	intergenic:VPS37D-DNAJC30/WBSCR22	4	0.26	chr7	73090050	73090072	+