

Table S1. Bacterial strains used in this study.

Strain	Species	Origin	<i>pirA/ pirB</i> genes	Reference
<i>Vp</i> _{AHPND}	<i>Vibrio parahaemolyticus</i>	Diseased shrimp (Taiwan)	+	[18]
<i>Vp</i> lab1	<i>V. parahaemolyticus</i>	Diseased shrimp (Ecuador)	+	Lab collection (not published)
<i>Vp</i> lab2	<i>V. parahaemolyticus</i>	Diseased shrimp (Vietnam)	+	Lab collection (not published)
CECT 8407	<i>V. parahaemolyticus</i>	Diseased shrimp (Mexico)	-	[64]
<i>Valg</i> lab1	<i>V. alginolyticus</i>	Healthy European seabass	-	Lab collection (not published)
CECT 8408	<i>V. harveyi</i>	Lesion of shrimp (Mexico)	-	[65]
CECT 4999	<i>V. vulnificus</i>	Diseased eel (Spain)	-	[66]

Table S2. Primers tested for *Vibrio parahaemolyticus* qPCR optimization.

Target gene	Primer	Primer sequence (5' - 3')	Reference
<i>tlh</i>	Tlh_F3	GCTACTCGAAAGATGATCC	[67]
	Tlh_LB	TAGATTGAAACGAGA	
<i>tdh</i>	D3	CCACTACCACTCTCATATGC	[68]
	D5	GGTACTAAATGGCTGACATC	
<i>tlh</i>	Tlh-Fw	AACTTCTGCGCCGAAGAG	[69]
	Tlh-Rv	CGGTGGATGTCCAAACAAGGA	
<i>tdh</i>	Tdh-Fe	GTAAAGGTCTCTGACTTTGGAC	[67]
	Tdh-Rv	CTACAGAACATAGGAATGTTGAAG	
<i>trh</i>	Trh-Fw	CCATCAATACCTTTCTCTCC	[67]
	Trh-Rv	ACCGTCATATAGGCCTAAC	
<i>pirA</i>	PirA-Fw	TTGGACTGTCGAACCAAACG	[50]
	PirA-Rv	GCACCCCCATTGGTATTGAATG	
<i>tlh</i>	VPF	AACCGTGGCGTTCCAGAA	[49]
	VPR	CCGTCAAACGAATCAGTGCTT	

Table S3. Detection of *Vibrio parahaemolyticus* strains by qPCR assay comparing PirA-Fw/PirA-Rv and VPF/VPR primer pairs.

<i>V. parahaemolyticus</i> strain	CT (qPCR PirA-Fw/PirA-Rv)	CT (qPCR VPF/VPR)
<i>Vp</i> _{AHPND}	14.89	12.32
<i>Vp</i> lab1	17.82	14.81
<i>Vp</i> lab2	16.31	14.42
CECT 8407	34.56	15.44

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