

Supporting informations

Concentrated O/W Emulsion Stability of Non-Ionic Chitosan Oligomer Surfactants Modified by Epoxidized Fatty Chains at pH7: Influence of Emulsification Conditions

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Characterizations of synthesized molecules

Characterization of COS-oxime

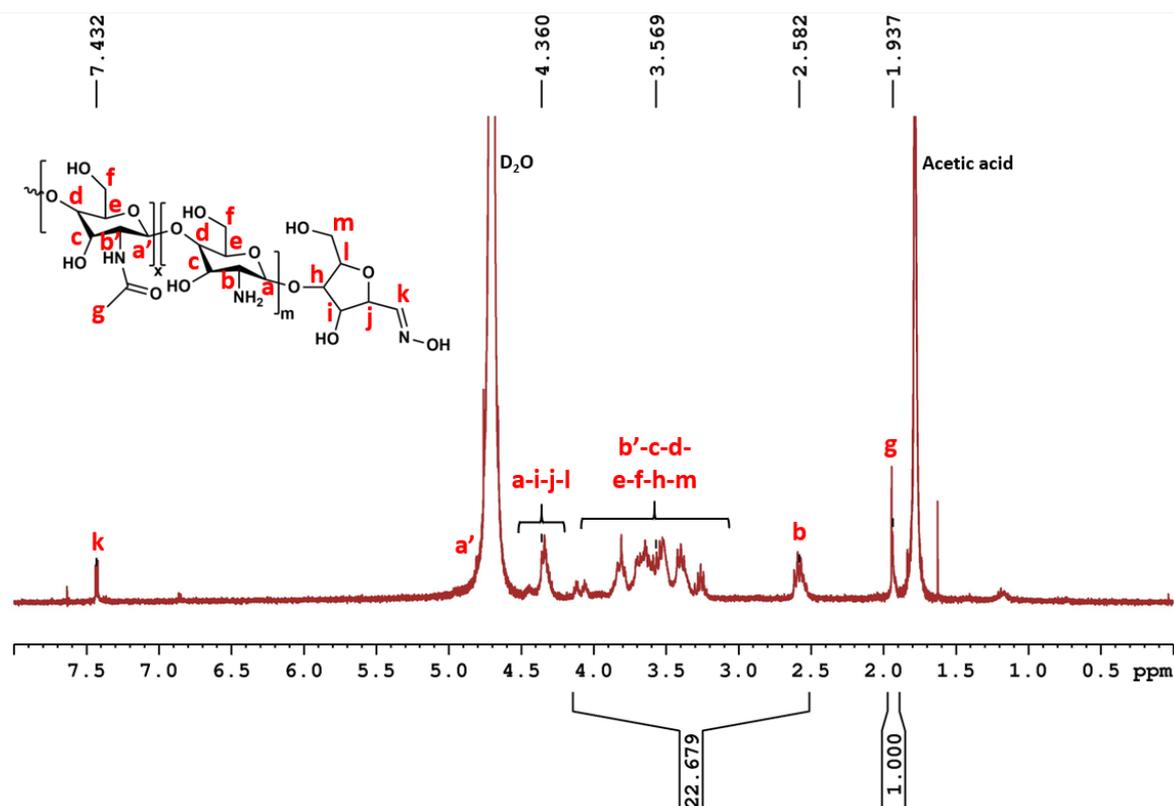


Figure S1. ¹H NMR (400 MHz, D₂O) spectrum, and proton assignments for DP10-oxime.

Characterization of COS-C9

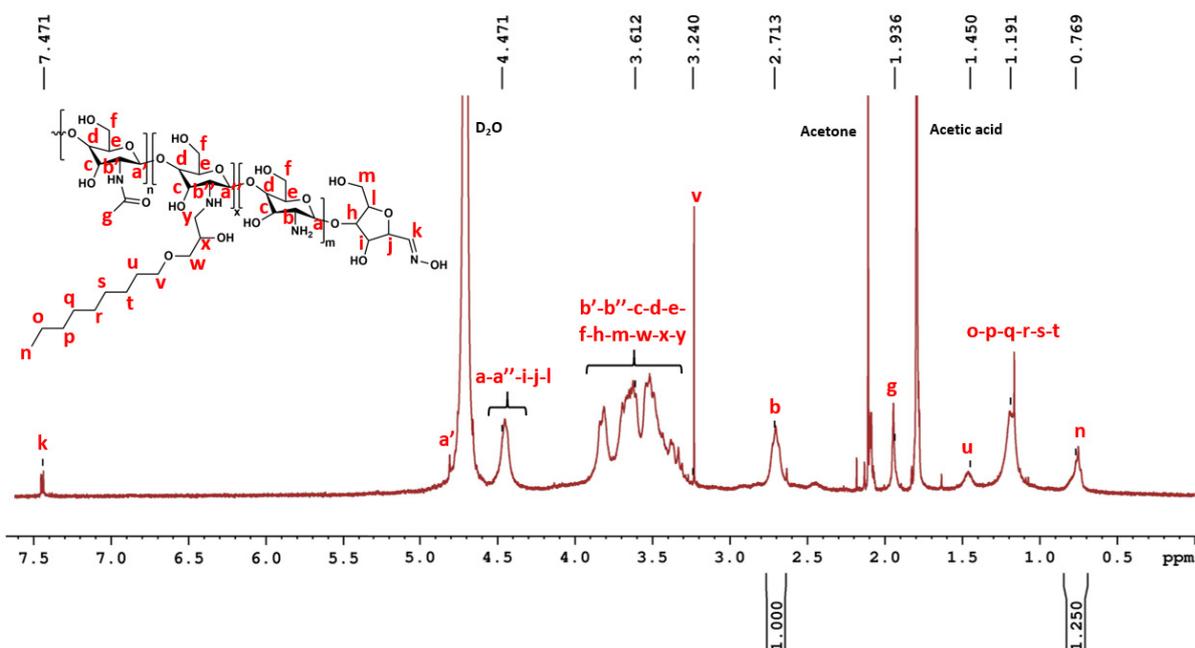


Figure S2. ^1H NMR (400 MHz, D_2O) spectrum, and proton assignments for DP10-C9.

Characterization of COS-C16

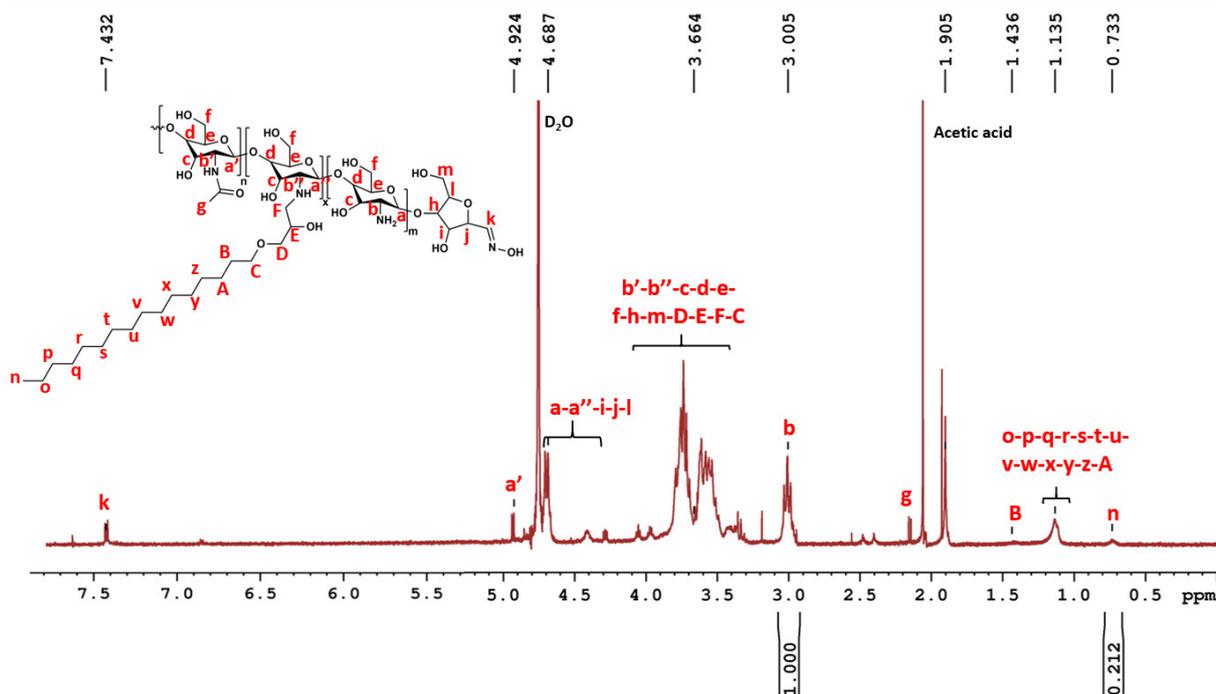


Figure S3. ^1H NMR (400 MHz, D_2O) spectrum, and proton assignments for DP10-C16.

Characterization of COS-card

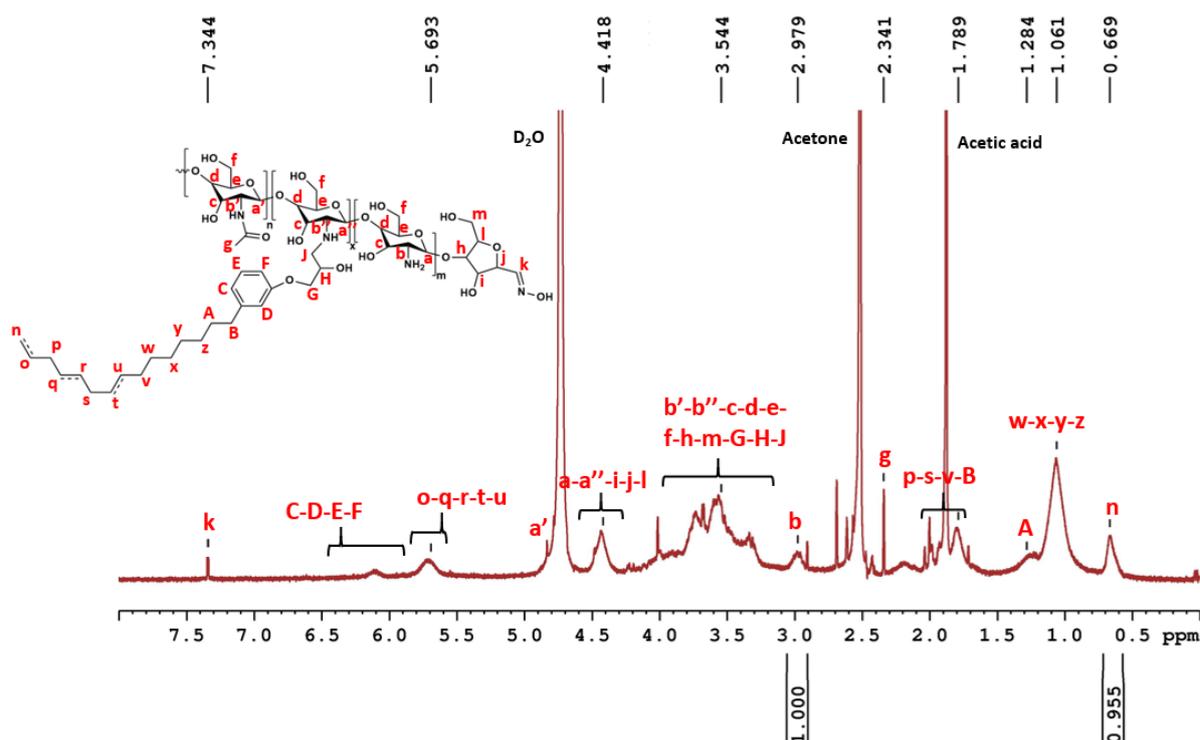


Figure S4. ^1H NMR (400 MHz, D_2O) spectrum, and proton assignments for DP10-card.

Characterizations of emulsions

Comparison of average droplet size using two different methods

| Method used | 1 minute | 2 minutes | 3 minutes | 4 minutes |
|------------------|----------|-----------|-----------|-----------|
| Light scattering | 14.9 | 14.2 | 13.9 | 13.9 |
| Image J | 14.8 | 13.9 | 13.7 | 13.8 |

Figure S5. Comparison of average droplet size (in μm) determination by light scattering and Image J (microcopy images) for the experiment as a function of stirring time. For the Image J results, average droplet size was calculated by averaging 200 droplets for each sample.

Rheological measurements to highlight the anti-thixotropic behavior

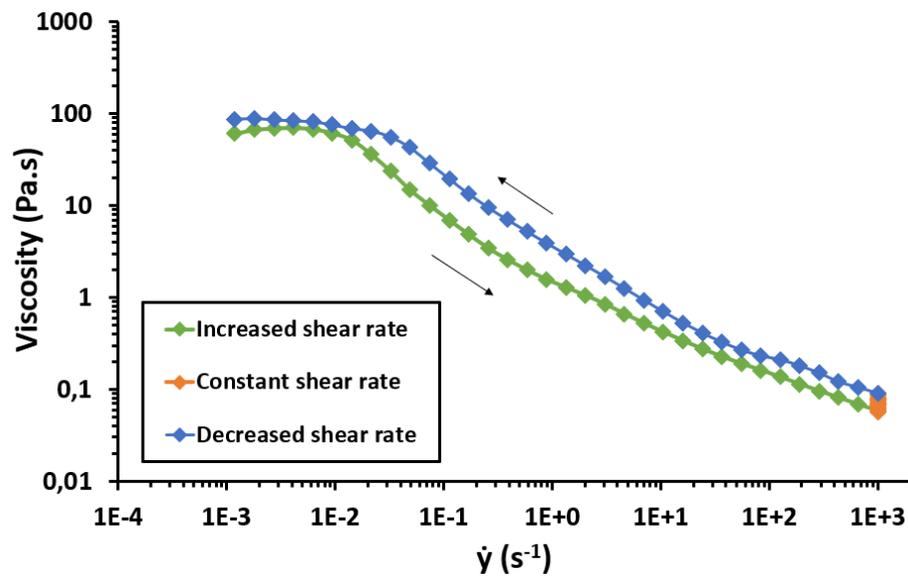


Figure S6. Viscosity profile of the emulsion with D10-card surfactant (stirring speed of 16500 rpm and stirring time of 4 minutes). An increase and then a decrease in shear rate were used to highlight the anti-thixotropic behavior.

Size distribution of emulsions as a function of stirring time

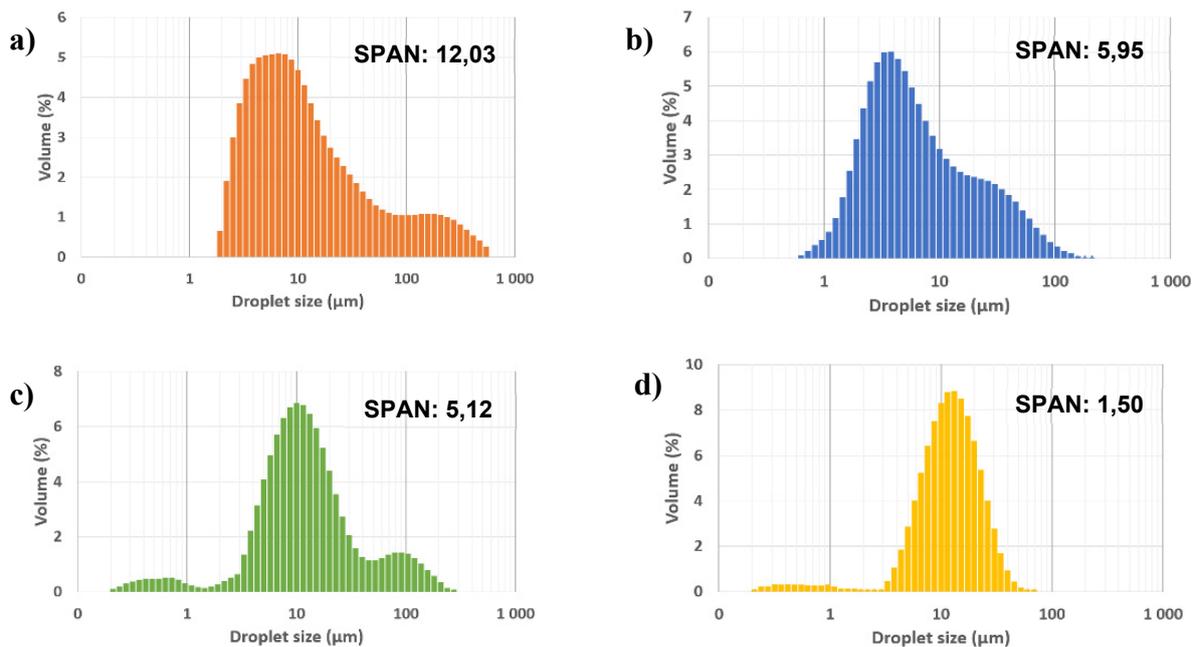


Figure S7. Size distribution of emulsion with DP10-card surfactant at a) 1 minute of stirring; b) 2 minutes of stirring; c) 3 minutes of stirring; d) 4 minutes of stirring.

Size distribution of emulsions as a function of stirring speed

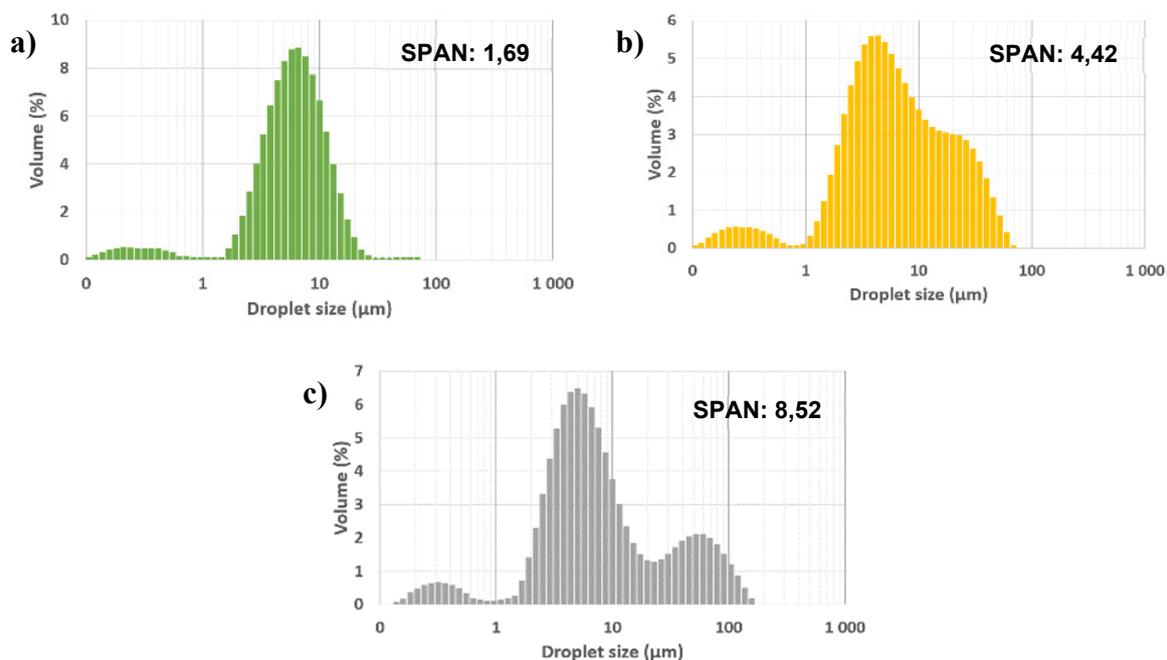


Figure S8. Size distribution of emulsion with DP10-card surfactant at a) 16500 rpm; b) 20500 rpm; c) 24500 rpm.

Highlighting the appearance of internal droplets by optical microscopy image

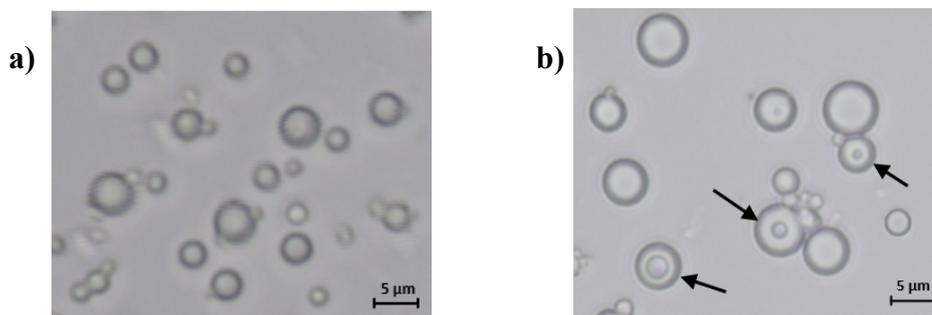


Figure S9. Optical microscopy images of a) an emulsion at a stirring speed of 16500 rpm; b) an emulsion at a stirring speed of 24500 rpm. Internal droplets are shown with arrows.

Highlighting the appearance of flocs by optical microscopy image

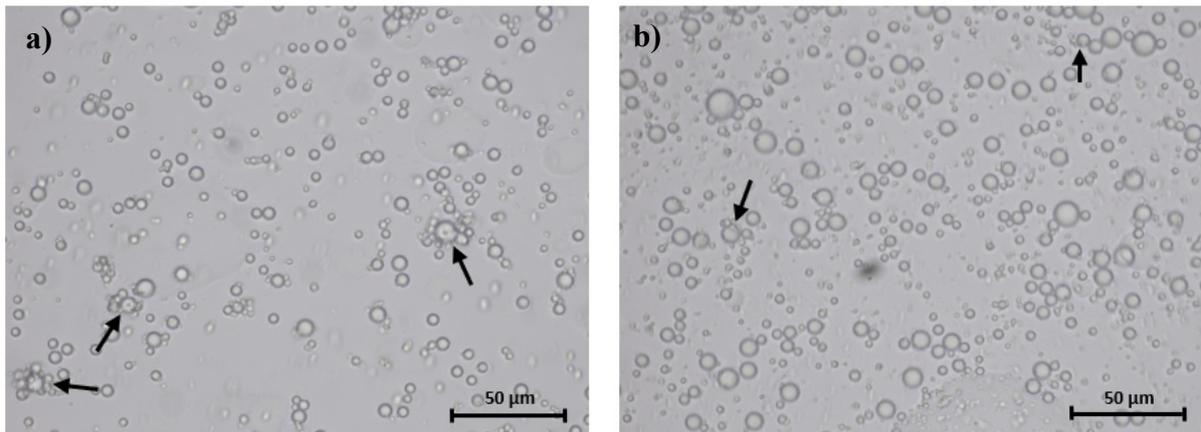


Figure S10. Optical microscopy images of diluted emulsions (x200) at t_0 with a) DP10-card surfactant; b) DP20-C9 surfactant. Flocs are shown with arrows.

%BS and %T evolution over time

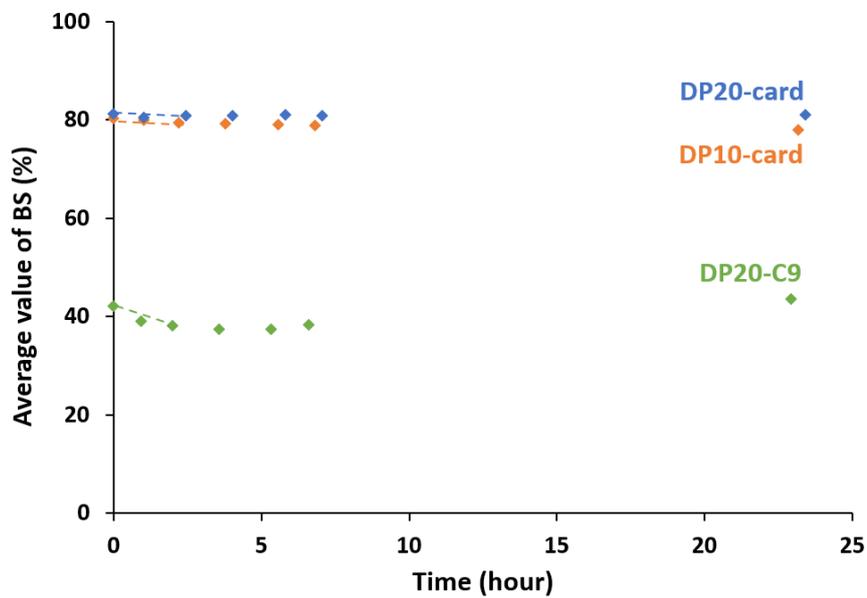


Figure S11. Backscattering percentage values over time for DP10-card, DP20-card and DP20-C9. Kinetics of flocculation/coalescence were measured during the first 4 hours of storage.

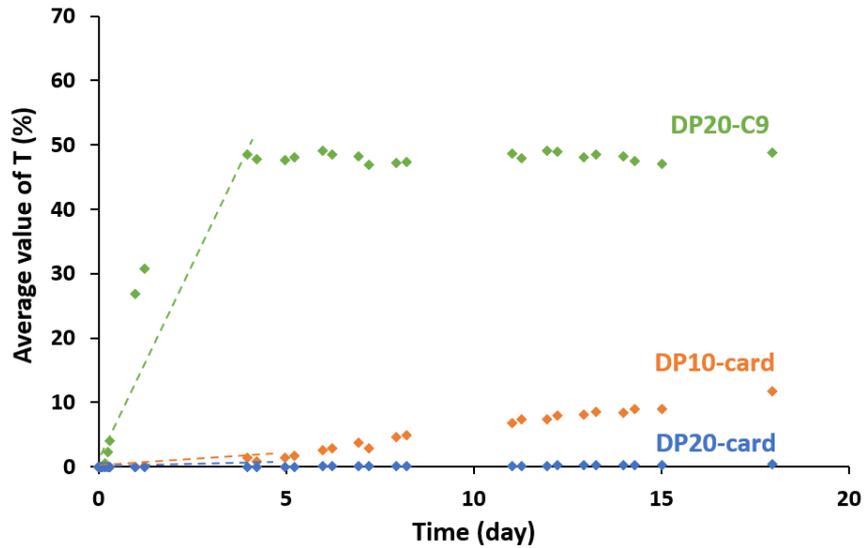


Figure S12. Transmission percentage values over time for DP10-card, DP20-card and DP20-C9. Kinetics of aqueous phase formation were measured during the first 5 days of storage using the slope at the origin.

Profiles evolution of Turbiscan

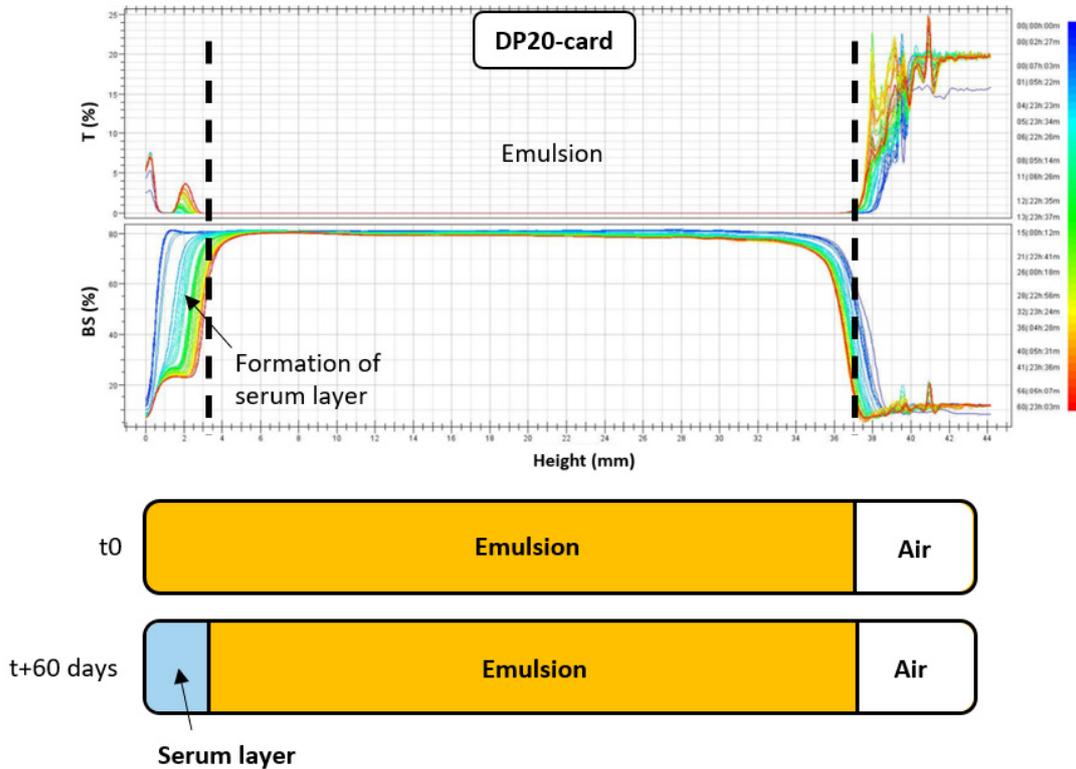


Figure S13. Transmission and backscattering profiles evolution as a function of time for the emulsion with DP20-card during 60 days. Horizontal representation of emulsion column at t0 and after 60 days.

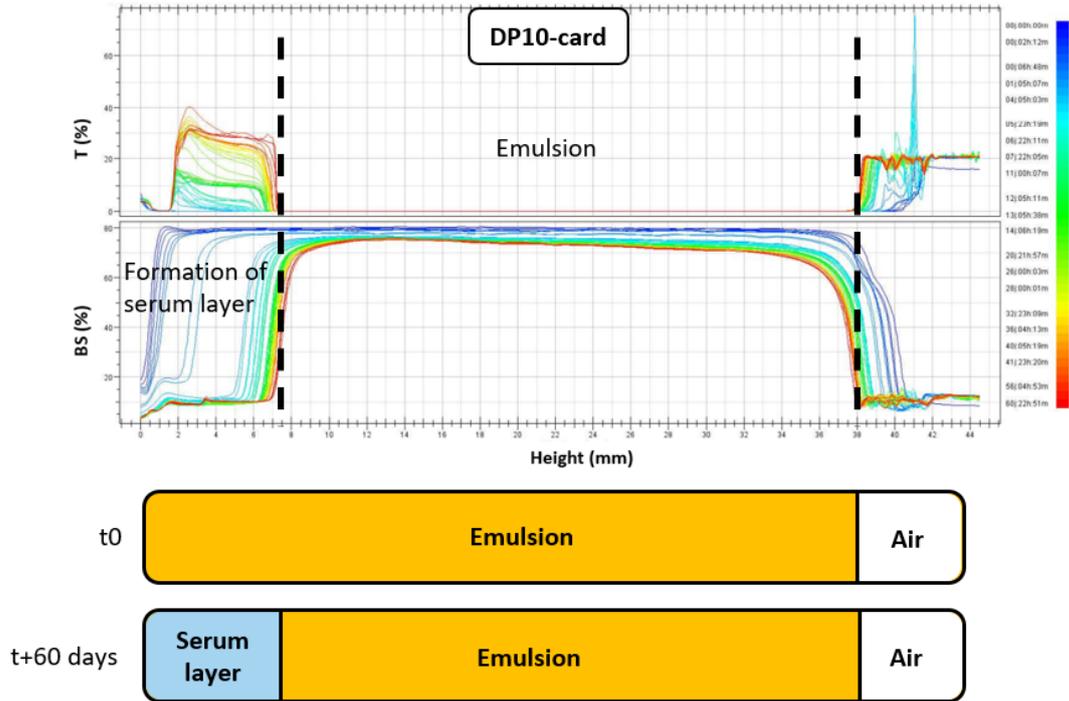


Figure S14. Transmission and backscattering profiles evolution as a function of time for the emulsion with DP10-card during 60 days. Horizontal representation of emulsion column at t_0 and after 60 days.

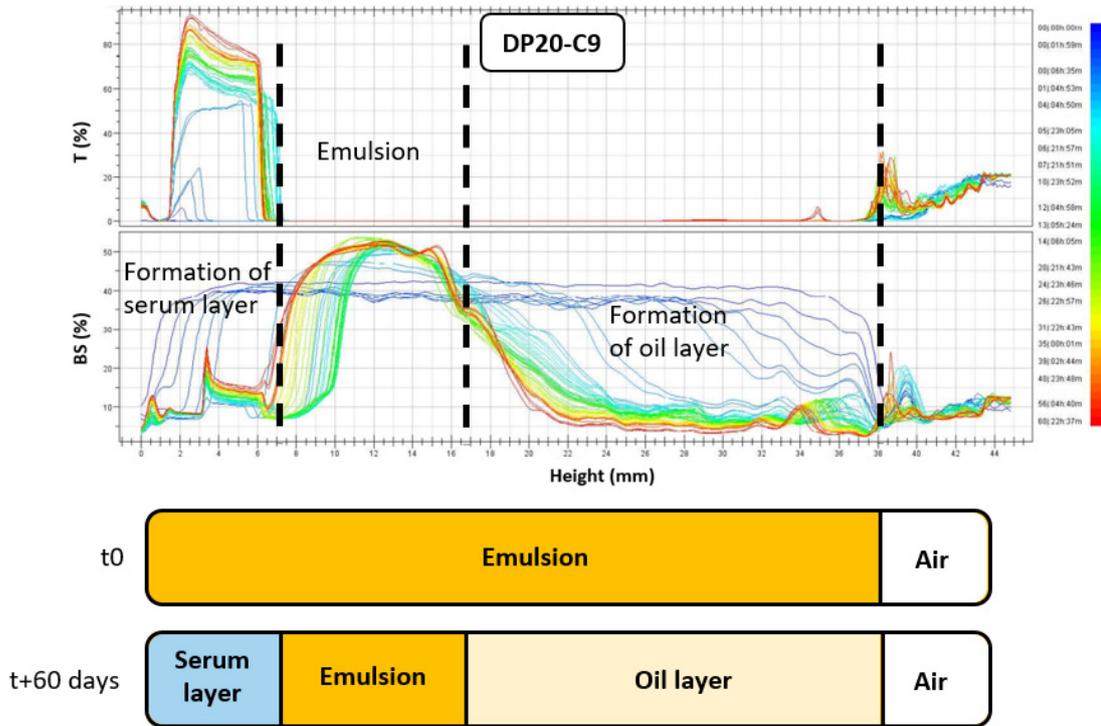


Figure S15. Transmission and backscattering profiles evolution as a function of time for the emulsion with DP20-C9 during 60 days. Horizontal representation of emulsion column at t_0 and after 60 days.