

**Table S1. Chromatographic conditions tested**

No	Stationary phase	Mobile phase	Volume composition of mobile phase
1	RP18W do HPTLC	acetonitrile + buffer pH=5.0	22.5:77.5
2	Silica gel 60F <sub>254</sub>	acetone + chloroform + ammonia	10 : 40: 0.5
3	Silica gel 60F <sub>254</sub>	n-hexane + acetone + ammonia	25 : 25: 0.5
4	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + methanol + acetic acid 80%	6:6:1:2:0.1
5	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + methanol + acetic acid 80%	6:6:2:2:0.1
6	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	6:6:6:2:0.2
7	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	18: 18: 7.5: 5.0: 0.3
8	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	3:3:3:2:0.1
9	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	6:6:2:2:0.1
10	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	1:3:3:2:0.1
11	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	3:1:3:2:0.2
12	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	3:3:3:2:0.1
13	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	6:6:2:2 :0.1
14	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	6:6:2.5:2:0.1
15	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	5.5:6:2.5:2:0.1
16	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	2:3:3:2:0.1
17	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	1:3:2:2:0.1
18	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	2:3:2:2:0.1
19	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	1:3:1.5:2:0.1

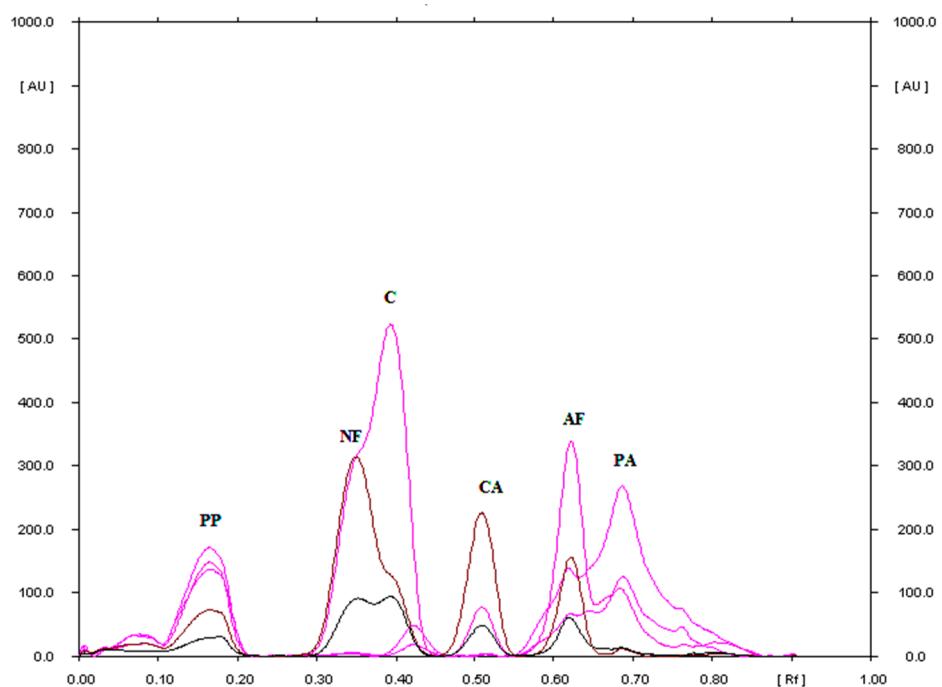
<b>20</b>	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	6:6:2.5:1.5:0.1
<b>21</b>	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + ethanol + acetic acid 80%	6:6:2:2:0.1
<b>22</b>	Silica gel 60F <sub>254</sub>	chloroform + toluene + ethyl acetate + methanol + glacial acetic acid	6:6:1:2:0.1

**Table S2.** The factors and their levels investigated in robustness test.

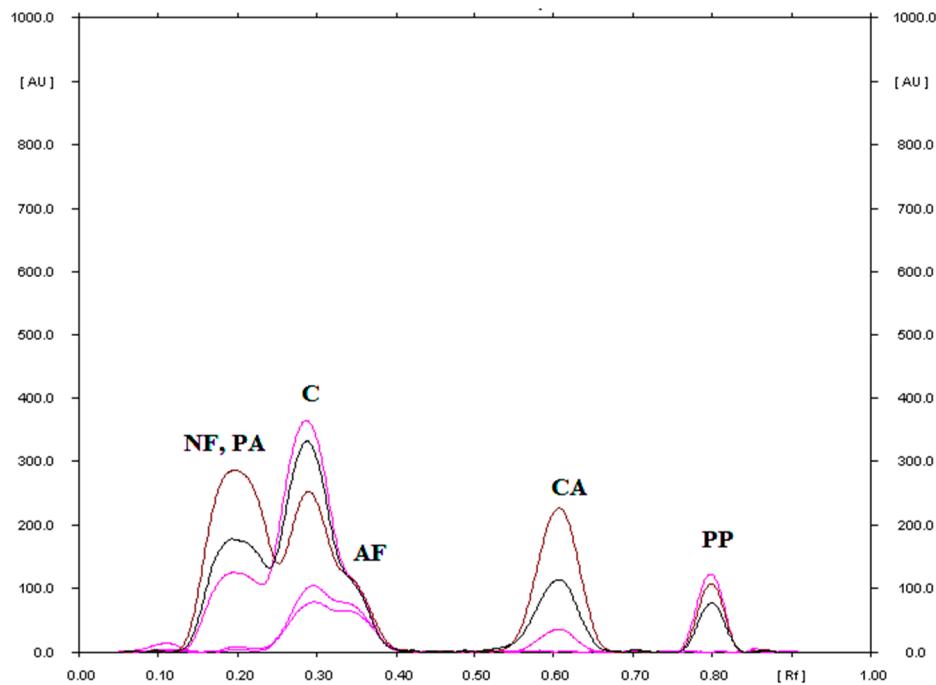
Symbol	Factors	Method Condition	Levels	
			+	-
X <sub>1</sub>	Temperature of plate activation [°C]	120	130	110
X <sub>2</sub>	Extraction time [min]	30	31	29
X <sub>3</sub>	Saturation time of the chamber [°C]	15	18	12
X <sub>4</sub>	Volume of chloroform [mL]	18.0	18.2	17.8
X <sub>5</sub>	Volume of toluene [mL]	18.0	18.2	17.8
X <sub>6</sub>	Volume of ethyl acetate [mL]	7.5	7.6	7.4
X <sub>7</sub>	Volume of ethanol [mL]	2.0	2.1	1.9

**Table S3** Experimental design matrix (2<sup>3</sup>) for robustness test.

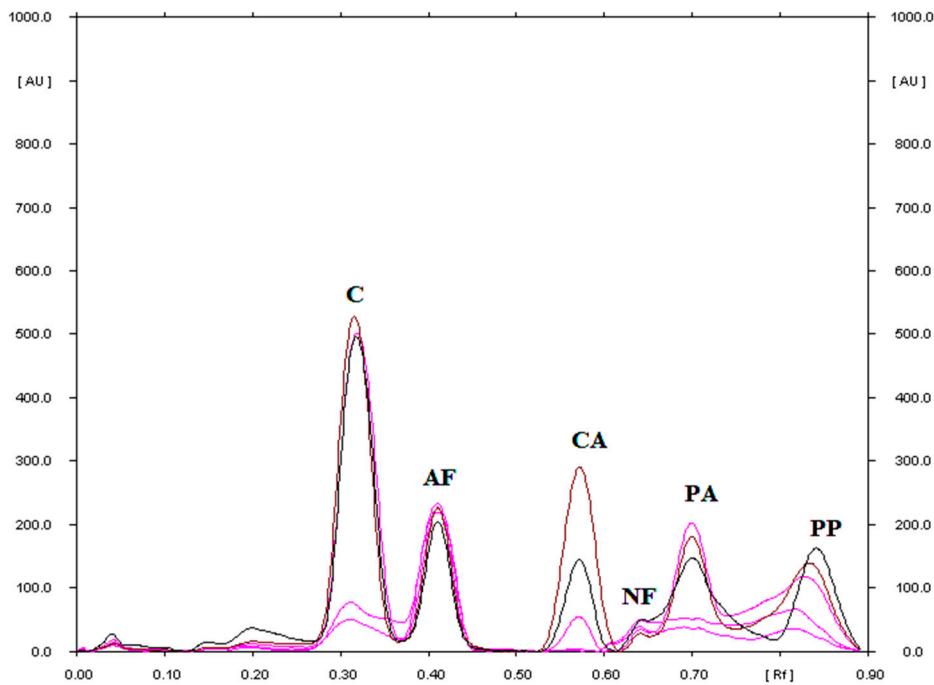
Experiment No	X <sub>1</sub>	X <sub>2</sub>	X <sub>3</sub>	X <sub>4</sub>	X <sub>5</sub>	X <sub>6</sub>	X <sub>7</sub>
1	+	+	+	+	+	+	+
2	+	+	-	+	-	-	-
3	+	-	+	-	-	+	-
4	+	-	-	-	+	-	+
5	-	+	+	-	+	-	-
6	-	+	-	-	-	+	+
7	-	-	+	+	-	-	+
8	-	-	-	+	+	+	-



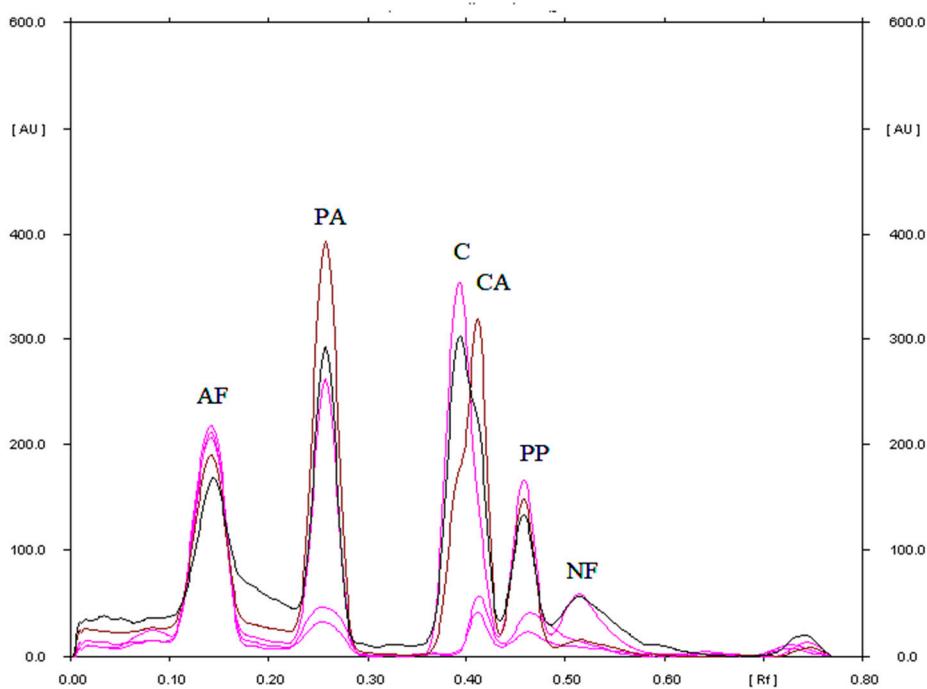
**Figure S1.** Densitogram of a mixture of standard substances: propyphenazone (PP), paracetamol (PA), caffeine (C), 4-chloroacetanilide (CA), 4-aminophenol (AF), and 4-nitrophenol (NF) made in the range of 250–350 nm, using RP18W plate and mobile phase: acetonitrile + buffer pH=5.0 (22.5:77.5, v/v).



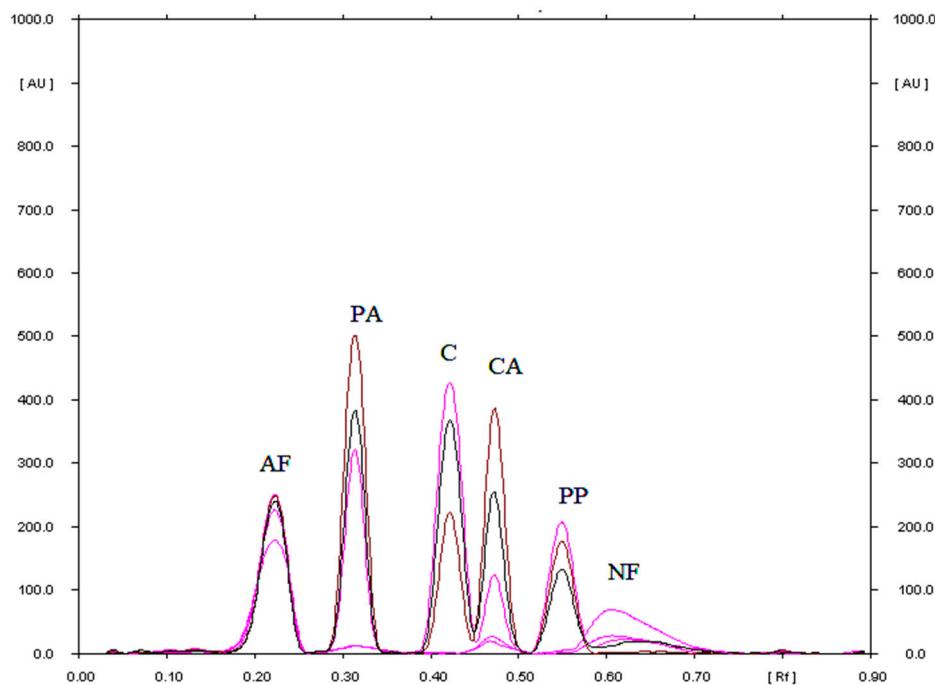
**Figure S2.** Densitogram of a mixture of standard substances: propyphenazone (PP), paracetamol (PA), caffeine (C), 4-chloroacetanilide (CA), 4-aminophenol (AF), and 4-nitrophenol (NF) made in the range of 250–350 nm, using silica gel 60F<sub>254</sub> plate and mobile phase: acetone + chloroform + ammonia (10 : 40: 0.5, v/v).



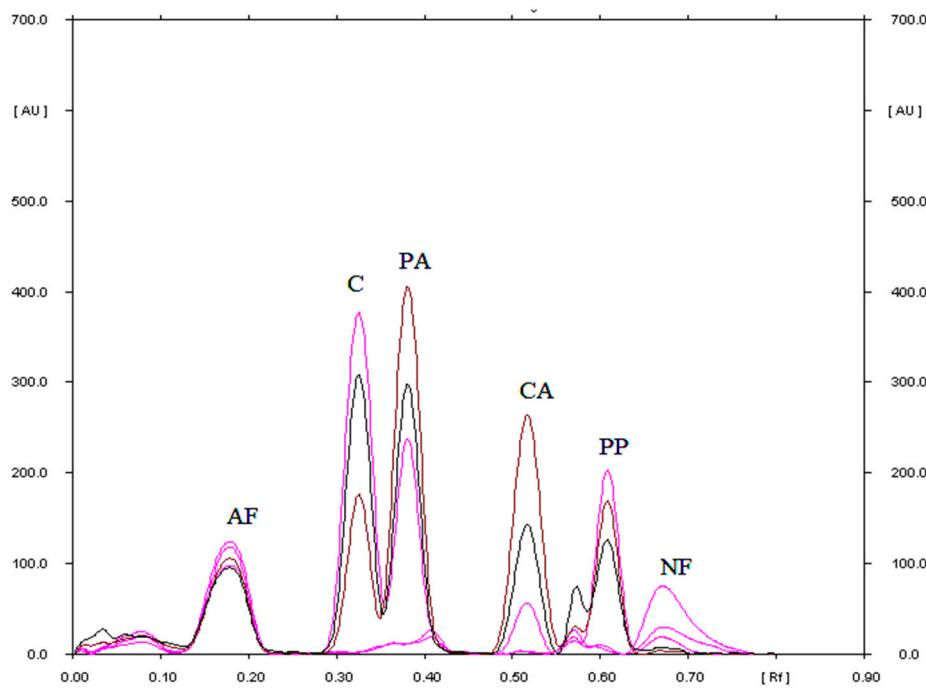
**Figure S3.** Densitogram of a mixture of standard substances: propyphenazone (PP), paracetamol (PA), caffeine (C), 4-chloroacetanilide (CA), 4-aminophenol (AF), and 4-nitrophenol (NF) made in the range of 250–350 nm, using silica gel 60F<sub>254</sub> plate and mobile phase: *n*-hexane + acetone + ammonia (25 : 25: 0.5, v/v).



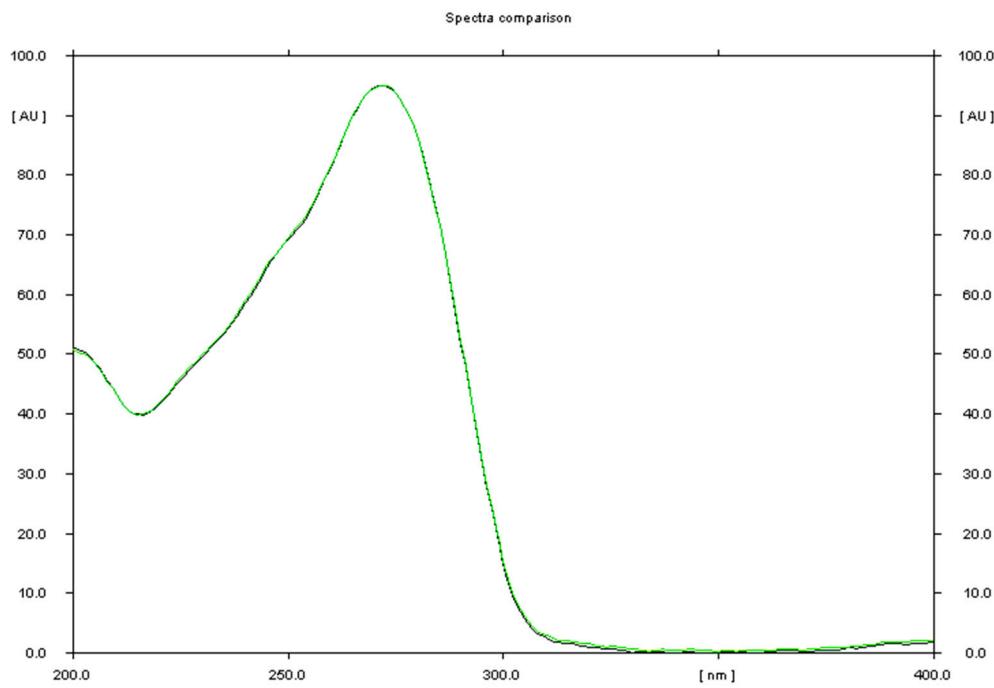
**Figure S4.** Densitogram of a mixture of standard substances: propyphenazone (PP), paracetamol (PA), caffeine (C), 4-chloroacetanilide (CA), 4-aminophenol (AF), and 4-nitrophenol (NF) made in the range of 250–350 nm, using silica gel 60F<sub>254</sub> plate and mobile phase: chloroform + toluene + ethyl acetate + methanol + acetic acid 80% (6:6:1:2:0.1, v/v).



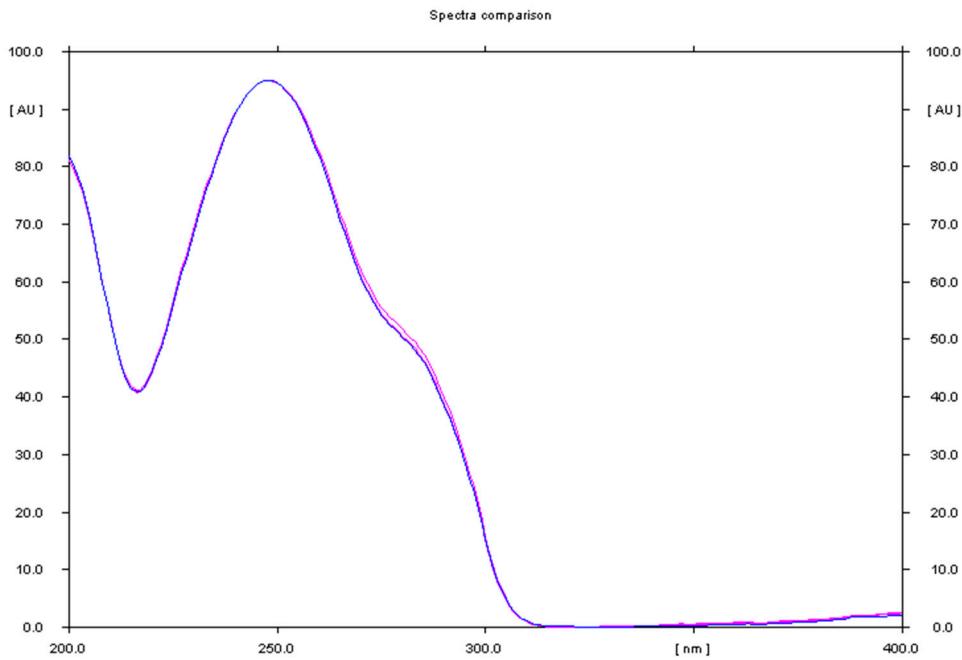
**Figure S5.** Densitogram of a mixture of standard substances: propyphenazone (PP), paracetamol (PA), caffeine (C), 4-chloroacetanilide (CA), 4-aminophenol (AF), and 4-nitrophenol (NF) made in the range of 250–350 nm, using silica gel 60F<sub>254</sub> plate and mobile phase: chloroform + toluene + ethyl acetate + methanol + acetic acid 80% (6:6:2:2:0.1, v/v).



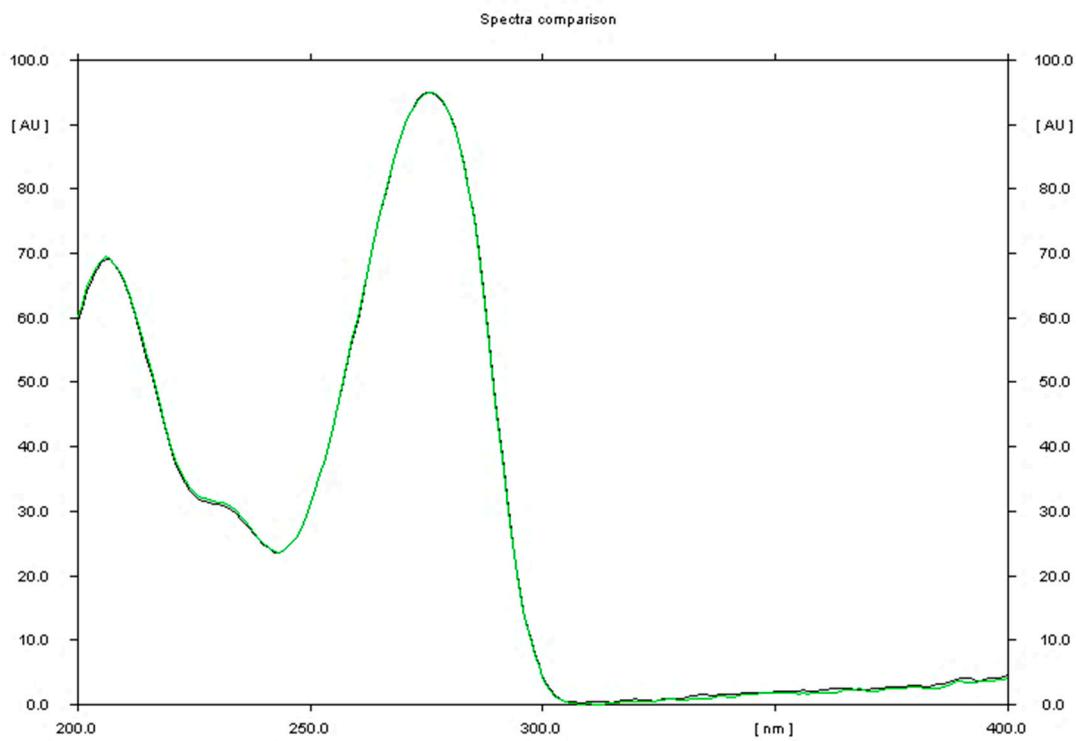
**Figure S6.** Densitogram of a mixture of standard substances: propyphenazone (PP), paracetamol (PA), caffeine (C), 4-chloroacetanilide (CA), 4-aminophenol (AF), and 4-nitrophenol (NF) made in the range of 250–350 nm, using silica gel 60F<sub>254</sub> plate and mobile phase: chloroform + toluene + ethyl acetate + ethanol + acetic acid 80% (6:6:6:2:0.2, v/v).



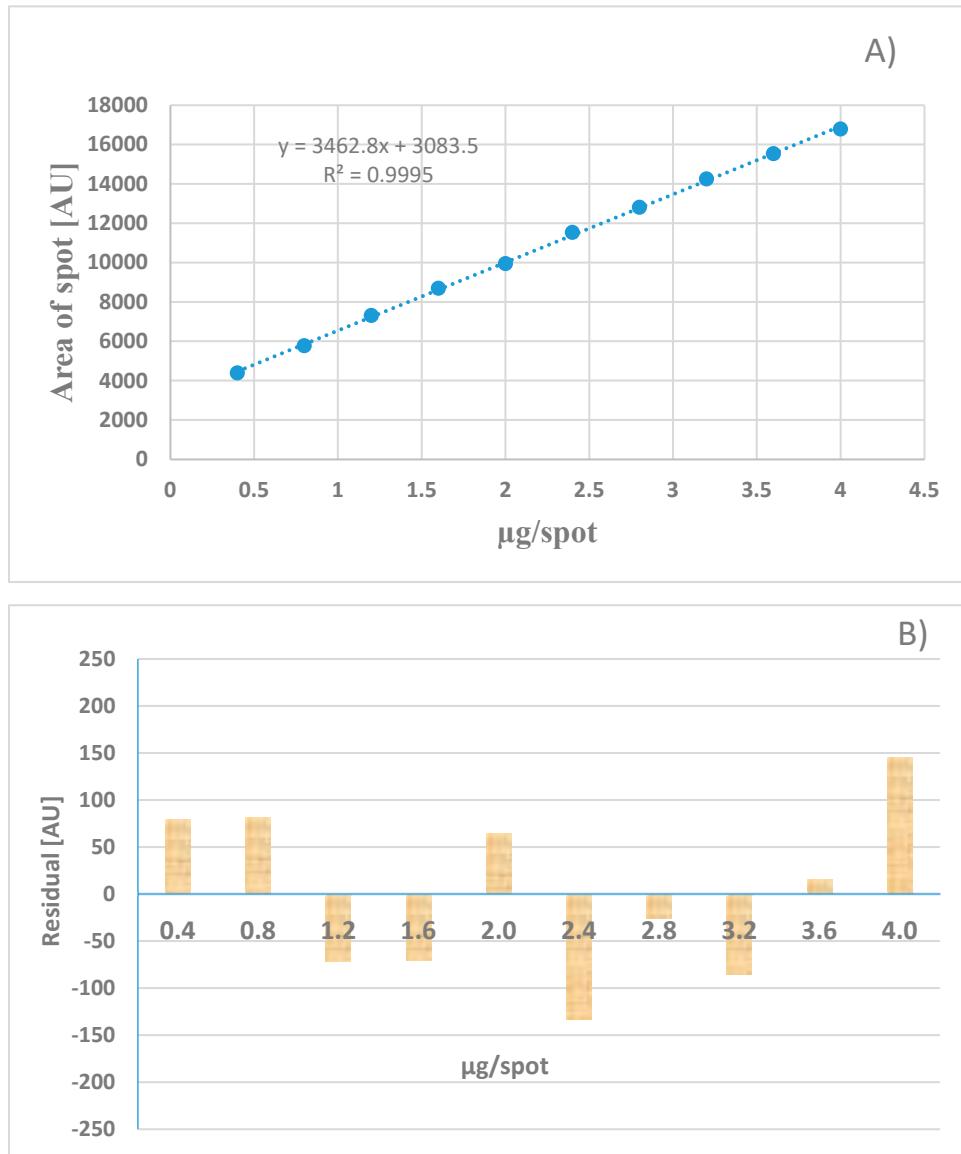
**Figure S7.** Comparison of the spectrodensitogram obtained for the standard substance propyphenazone with the spectrodensitogram obtained for propyphenazone, the source of which was sample of Saridon.



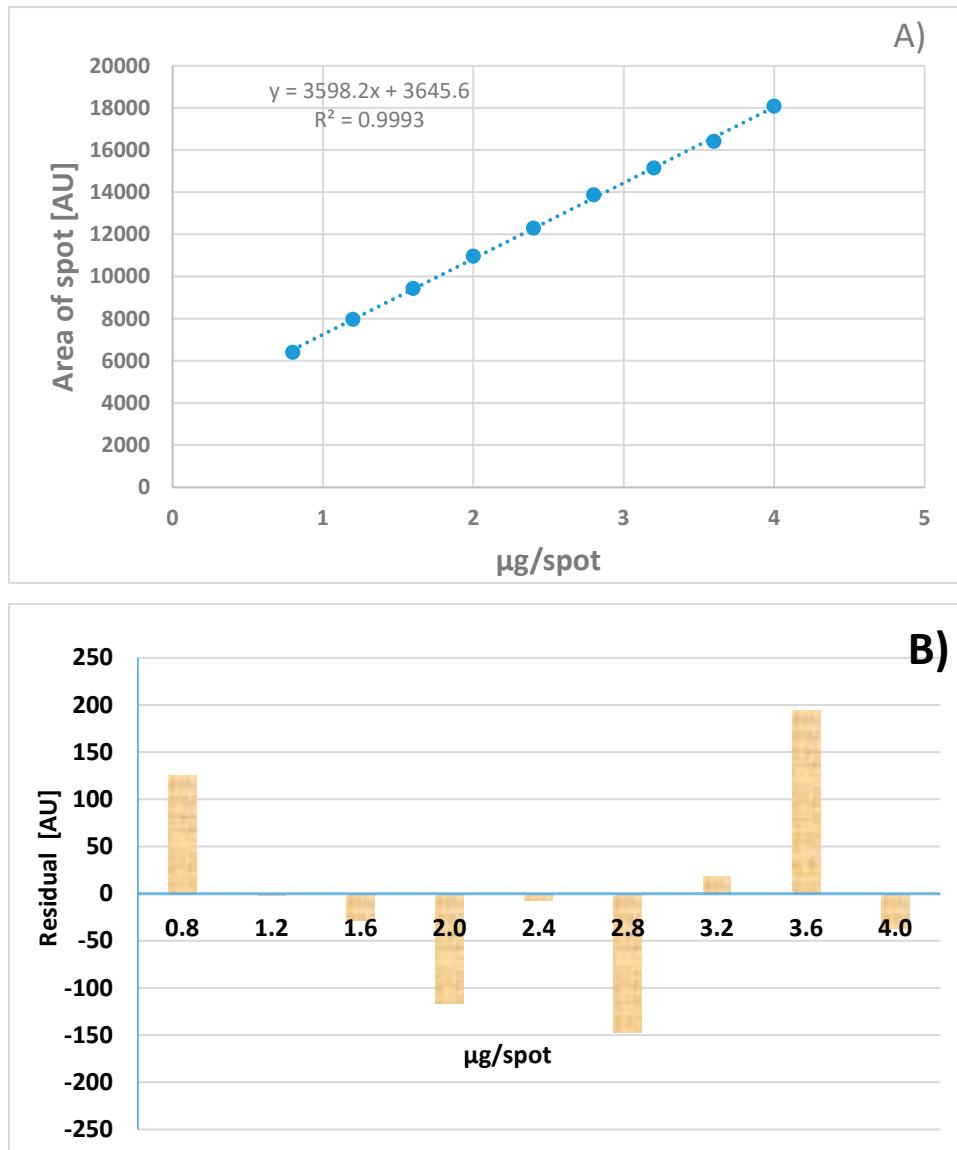
**Figure S8.** Comparison of the spectrodensitogram obtained for the standard substance paracetamol with the spectrodensitogram obtained for paracetamol, the source of which was sample of Saridon



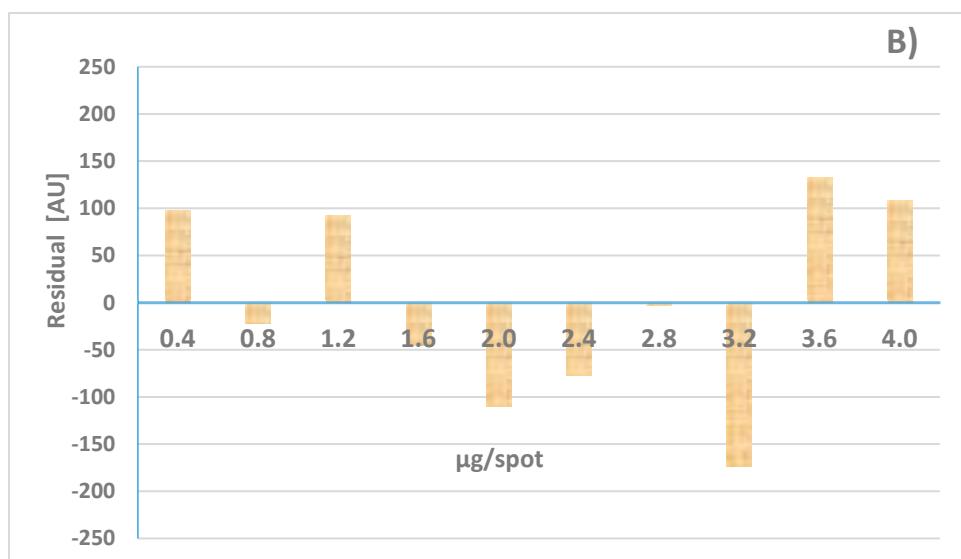
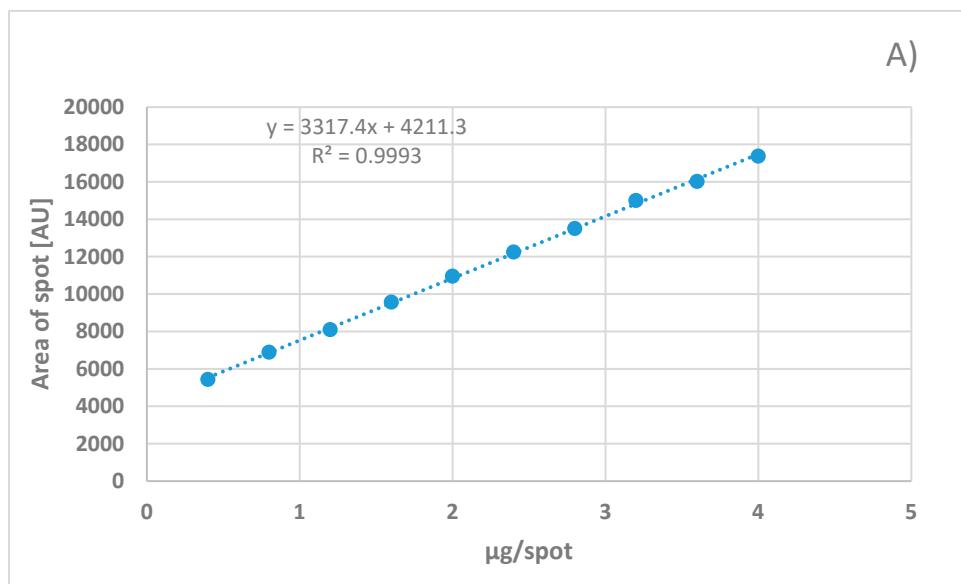
**Figure S9.** Comparison of the spectrodensitogram obtained for the standard substance caffeine with the spectrodensitogram obtained for caffeine, the source of which was sample of Saridon



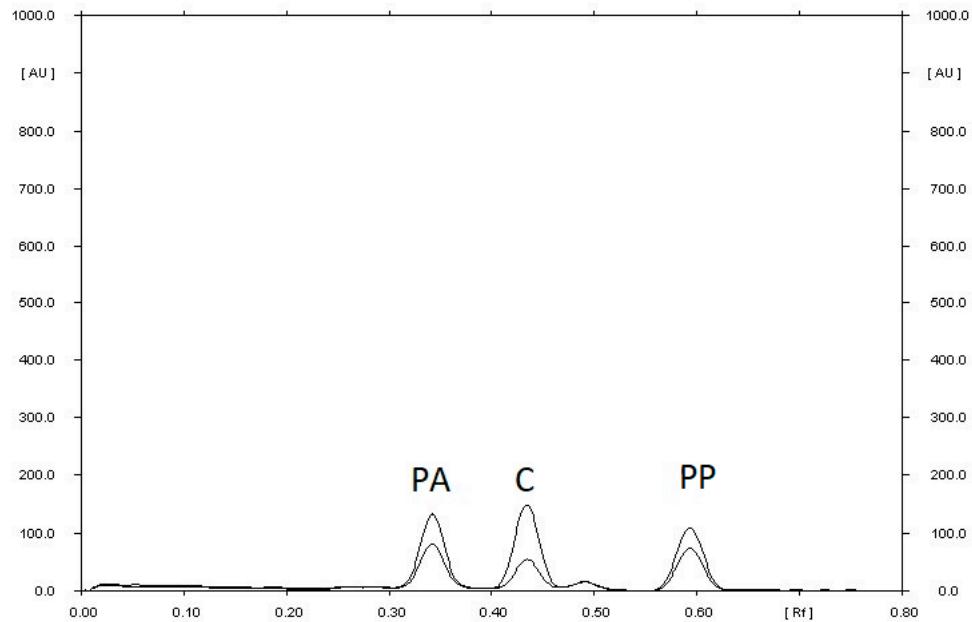
**Figure S10.** Calibration plot (A) and plot of residuals (B) for paracetamol (PA) in the linear working range mobile phase: chloroform + toluene + ethyl acetate + ethanol + acetic acid (80%) in a volume ratio of 18: 18: 7.5: 5.0: 0.3.



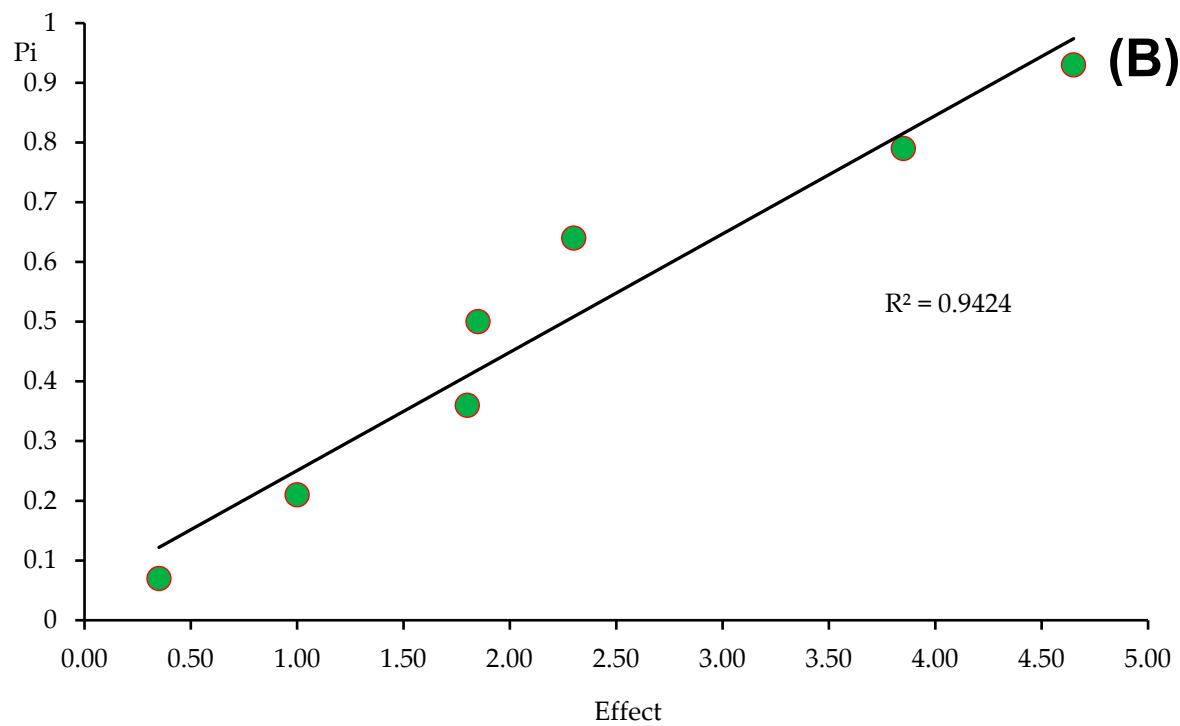
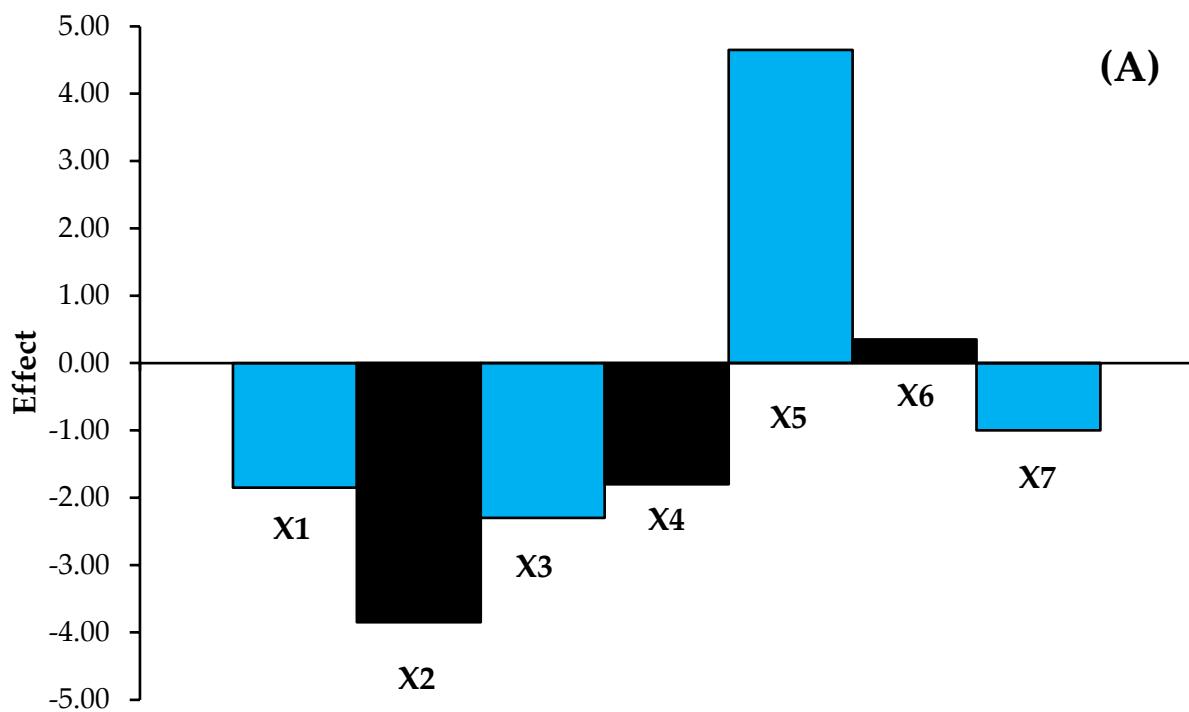
**Figure S11.** Calibration plot (A) and plot of residuals (B) for propyphenazone (PP) in the linear working range mobile phase: chloroform + toluene + ethyl acetate + ethanol + acetic acid (80%) in a volume ratio of 18: 18: 7.5: 5.0: 0.3.



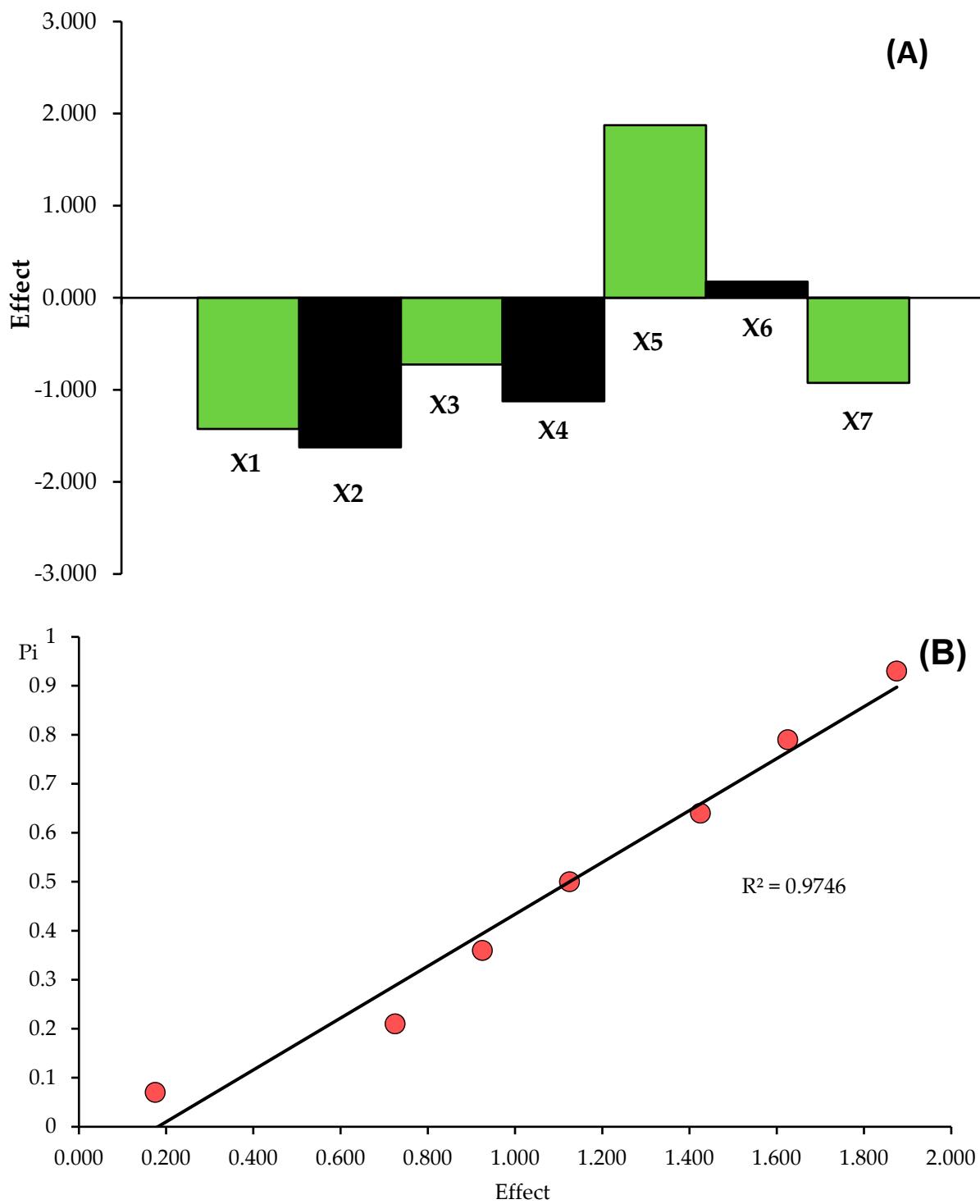
**Figure S12.** Calibration plot (A) and plot of residuals (B) for caffeine (C) in the linear working range mobile phase: chloroform + toluene + ethyl acetate + ethanol + acetic acid (80%) in a volume ratio of 18: 18: 7.5: 5.0: 0.3.



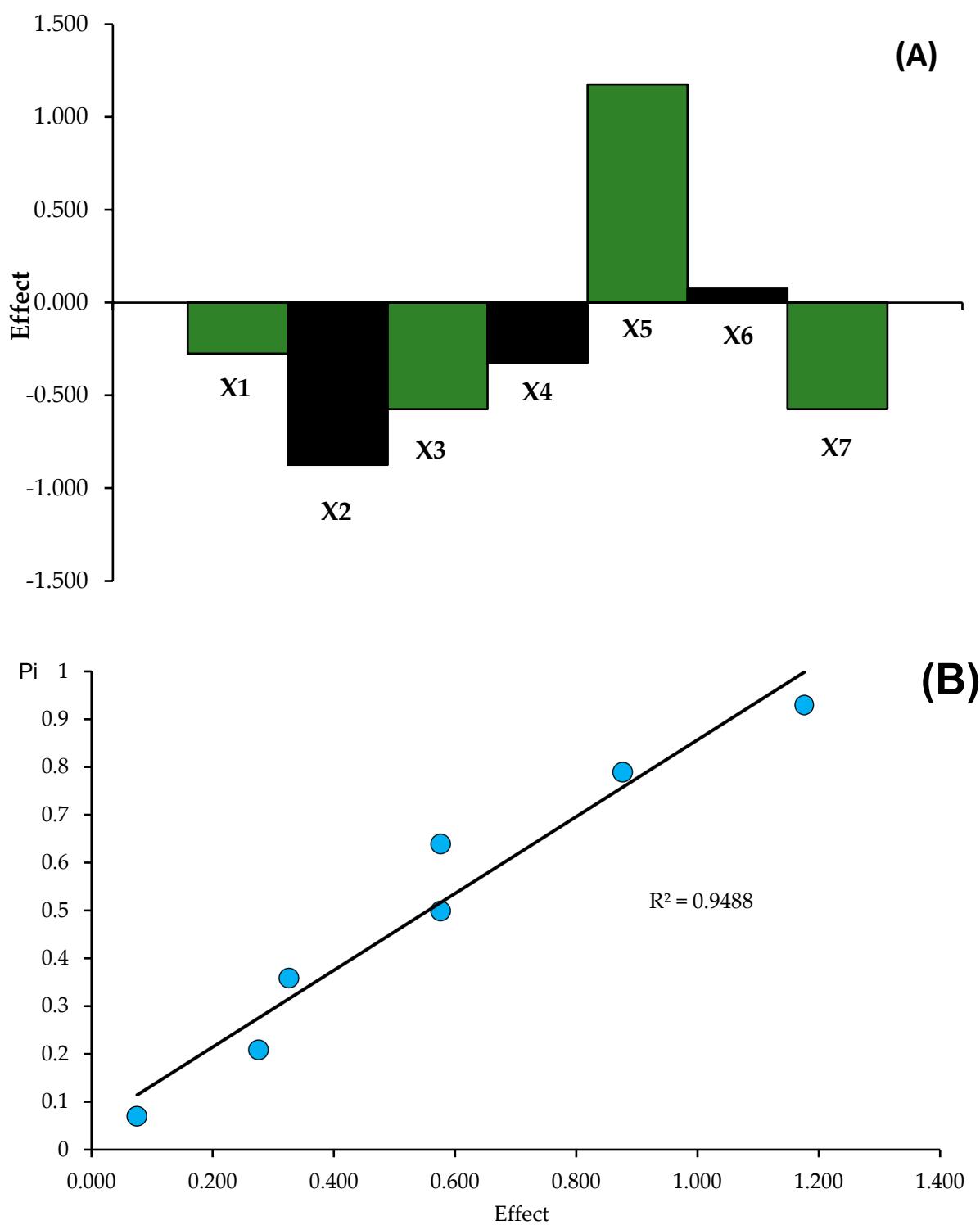
**Figure S13.** Densitogram of standard mixture of PA, C, PP (each standard about concentration 0.20  $\mu\text{g}/\text{spot}$ ).



**Figure S14.** Robustness test: the effects of factors (A), and half-normal probability plot of effects (B) for determination of paracetamol (PA) in Saridon tablets.



**Figure S15.** Robustness test: the effects of factors (A), and half-normal probability plot of effects (B) for determination of propyphenazone (PP) in Saridon tablets.



**Figure S16.** Robustness test: the effects of factors (A), and half-normal probability plot of effects (B) for determination of caffeine (C) in Saridon tablets.

**Table S4.** Literature values of LOD and LOQ of PA, PP, and C investigated by HPLC and micellar liquid chromatography techniques.

Method	Stationary phase	Mobile phase	LOD and LOQ [ μg/mL] of			Ref
			PA	PP	C	
RP-HPLC	Nucleosil C18	Water + methanol, 20:80, v/v)	LOD: 0.30 LOQ: 0.88	LOD: 0.25 LOQ: 0.41	LOD: 0.36 LOQ: 0.66	[20]
RP-HPLC	ODS column	Ortho phosphoric acid 0.1% + acetonitrile, 85:15, v/v	LOD: 0.18 LOQ: 0.55	LOD: .05 LOQ: 0.14	LOD: .03 LOQ: 0.09	[22]
RP-HPLC	Hypersil C18 BDS	0.2 mol/L Tetrabutylammonium bisulphate + methanol, 100:45, v/v	LOD: 2.20 LOQ: 4.60	LOD: 0.09 LOQ: 0.12	LOD: 0.42 LOQ: 0.62	[23]
RP-HPLC	C18 column	Acetonitrile+methanol+water (25:25:50 v/v)	LOD: 6.57 LOQ: 19.90	LOD = 5.19 LOQ = 15.74	LOD: 3.098 LOQ: 0.39	[24]
RP-HPLC	Chromolith RP-18e	Acetonitrile + water (30:70 v/v/)	LOD: 0.4 LOQ: 1.2	LOD: 0.7 LOQ: 2.4	LOD: 0.5 LOQ: 1.6	[25]
RP-HPLC	C18 column	Phosphate buffer (pH 9; 0.05 M) : methanol(80:20 v/v)	LOD: 0,3 LOQ: 1	-	LOD: 0,3 LOQ: 1	[13]
RP-HPLC	C18 column	Methanol+water, 40:60, v/v	LOD: 0.01 LOQ: 0.033	-	LOD: 0.005 LOQ: 0.016	[7]
RP-HPLC	Hypersil C18 BDS	0.02 M tetrabutylammonium bisulfate : methanol (100:45 v/v)	LOD: 2.20 LOQ: 4.40	-	LOD: 0.42 LOQ: 0.62	[6]
RP-HPLC-UV	μ-Bondapak C18	Methanol+water+triethylamine (60+40+0.1, v/v)	-	LOD: 5.0 LOQ: 8.0	LOD: 1.5 LOQ: 5.0	[17]
LC	Cyanopropyl column	Acetonitrile +12mM ammonium acetate(25,75 v/v)	-	LOD: 0.0065 LOQ: 0.022	LOD: 0.004 LOQ: 0.013	[15]
Micellar electrokinetic capillary liquid chromatograph						
	Silica capillary	20 mM borate buffer	LOD: 0.6 LOQ: 2.0	LOD: 0.8 LOQ: 3.0	LOD: 0.6 LOQ: 3.0	[31]
		y				

Micellar liquid chromatography	ODS C18	40mM sodium dodecyl sulphate + 10% propan-1-ol + 0,3% triethylamine in 0,02 M phosphoric acid	-	-	LOD: 0.6 LOQ: 0.8	[42]
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